In vitro systems for assessing vaccine quality and elucidating vaccine-induced immune mechanisms

Aurora Signorazzi¹, Marilena Etna², Gabriela Tapia Calle¹, Shuran Gong¹, Eliana Coccia² and Anke Huckriede¹

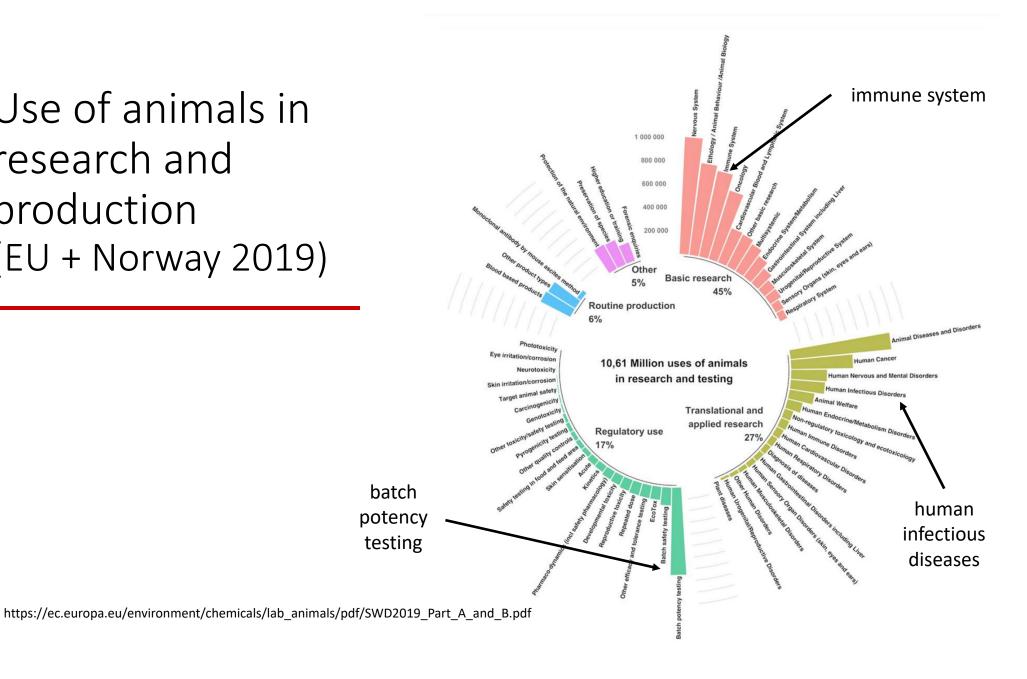
¹ Department of Medical Microbiology & Infection Prevention, University Medical Center Groningen and University of Groningen, Groningen, The Netherlands

² Department of Infectious Diseases, Istituto Superiore di Sanitá, Rome, Italy

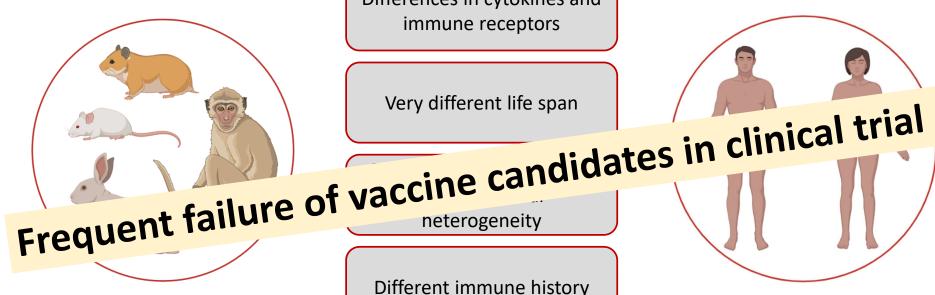




Use of animals in research and production (EU + Norway 2019)



Animals are not small humans



Differences in cytokines and immune receptors

Different immune history

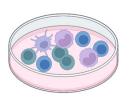


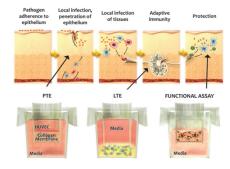




Modeling the human immune system in vitro

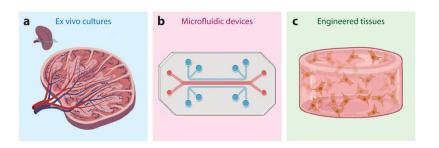
- Immune response induction
 - Blood cell-based models
 - Artificial lymph nodes
 - MIMIC®





Drake III et al Disruptive Sci & Techn 2012

- Immune response effects
 - Organ slices
 - Organ-on-a-chip
 - Organoids

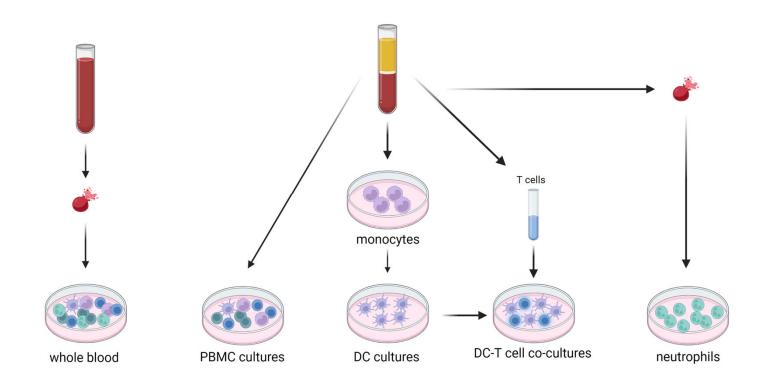


Hammel et al Ann Rev Biomed Eng 2021





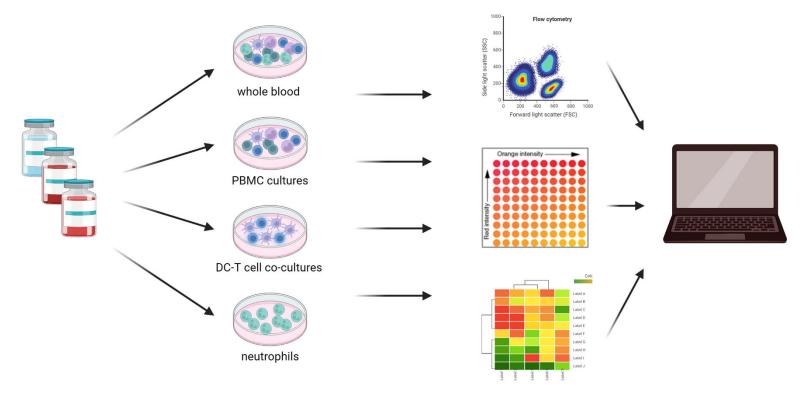
Simple blood cell-based systems







Experimental setup







Example 1: Use of PBMCs for vaccine comparison





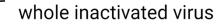
The model: Different types of influenza vaccines



 High immunogenicity in naïve individuals



- Th1-like response
- High IgG2a
- High levels of IFNγproducing T cells





split



subunit



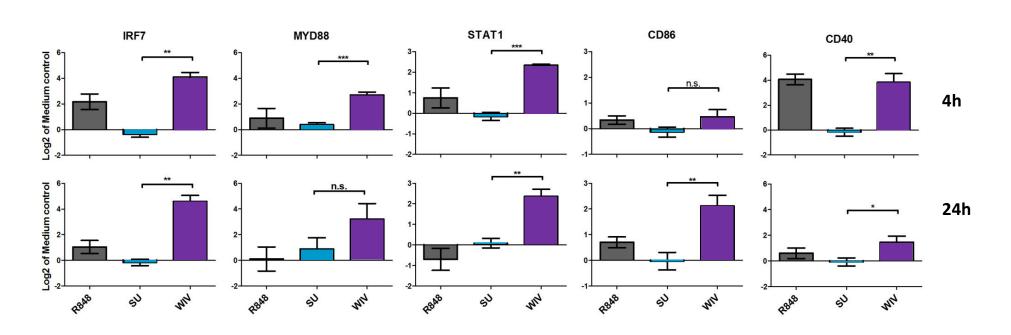
 'Low' immunogenicity in naïve individuals



- Th2-like response
- · High IgG1
- High levels of IL4producing T cells



WIV but not SU vaccine activates distinct immune related gene expression pathways in DCs

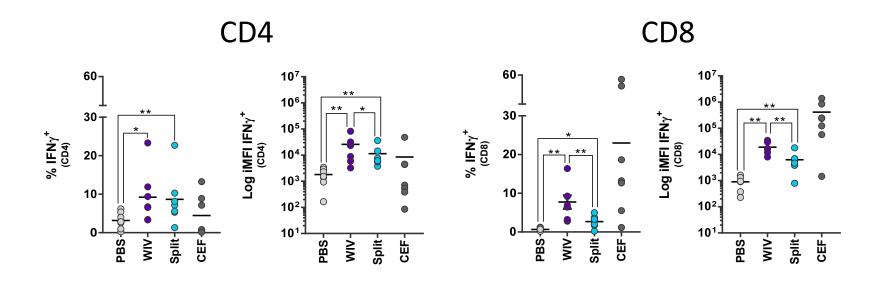


Note: response to R848 and WIV similar but not identical!





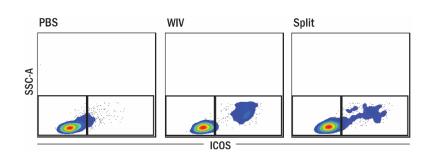
WIV is superior to split vaccine in inducing IFNy production in CD4 and CD8 T cells

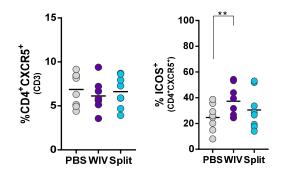


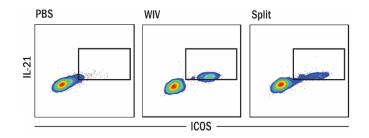


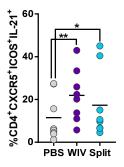


WIV and split vaccine stimulate T_{FH} cells which are associated with antibody responses













Example 2:

Use of PBMCs in vaccine batch quality control for Tick Borne Encephalitis (TBE) vaccine







Current approach in TBE vaccine quality control

- Vaccine: Formaldehyde-inactivated virus adjuvanted with Alum
- Vaccine batch assessment:
 - Physico-chemical characterization
 - Mandatory animal challenge experiments to prove and measure vaccine potency
- Problems
 - Large number of animals required
 - High level of discomfort for animals
 - Large variation in outcome of challenge experiments (accepted confidence interval 30-300%)





Alternative approach for TBE vaccine quality control

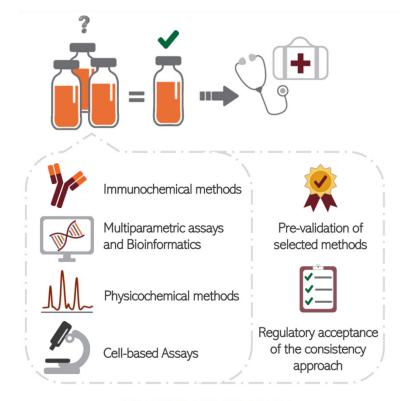
 Batch-to-batch consistency approach











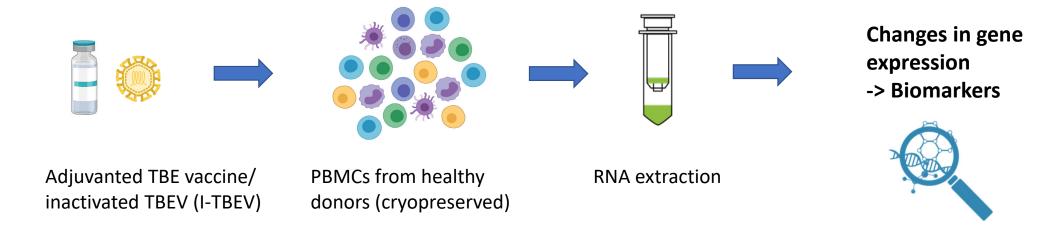
CONSISTENCY APPROACH





Quality assessment of tick-borne encephalitis(TBE) vaccine batches

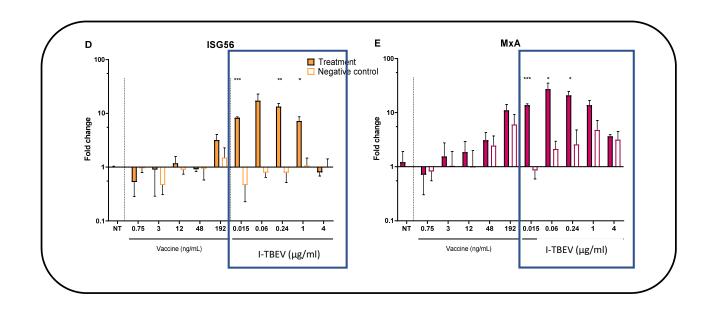
Aim: Identify biomarkers for quality/potency of vaccine







Human PBMCs respond to inactivated-TBEV (I-TBEV) by upregulation of innate immune responses







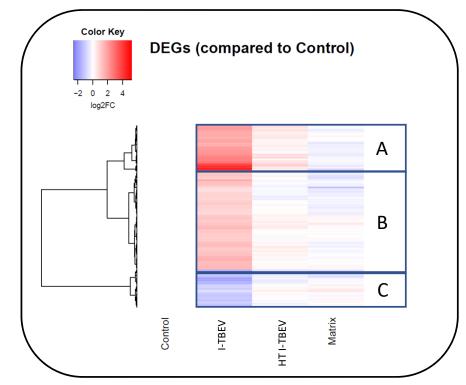
RNAseq identifies antiviral defense and IFN signaling as I-TBEV-induced pathways

3 clusters:

A. Genes strongly upregulated in I-TBEV-stimulated cells
[ISG56, MxA, Viperin, MDA5, CCL8, CXCL10] ->

antiviral defense, IFN signalling

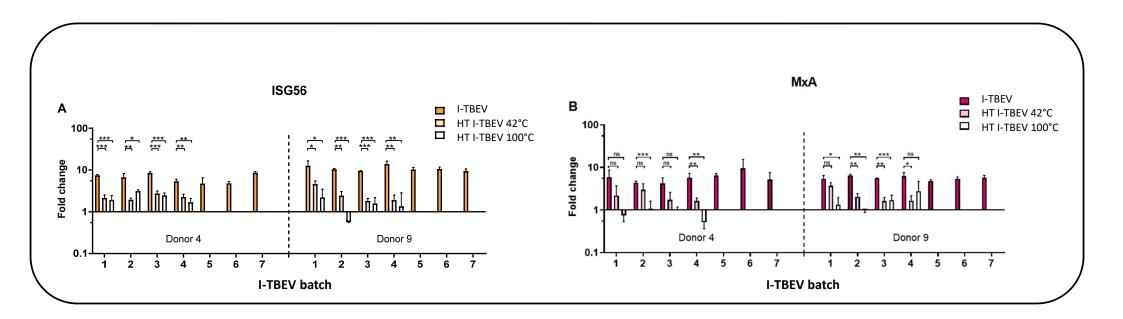
- B. Genes upregulated in I-TBEV-stimulated cells [selected chemokines/cytokines] -> immunity
- C. Genes downregulated in I-TBEV-stimulated cells [ribosomal proteins; lipid metabolism; ICAM-1, DC-SIGN, MHC-II, CD14]







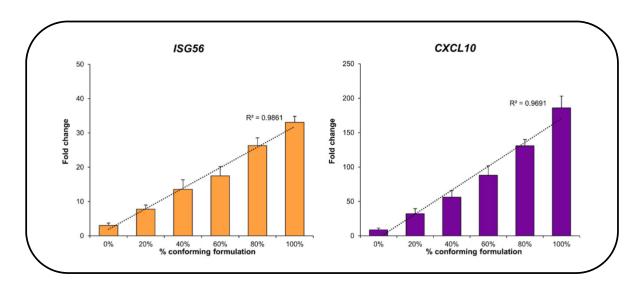
PBMCs respond to different batches of I-TBEV in a consistent way







PBMCs detect the amount of conforming I-TBEV in a batch with high resolution

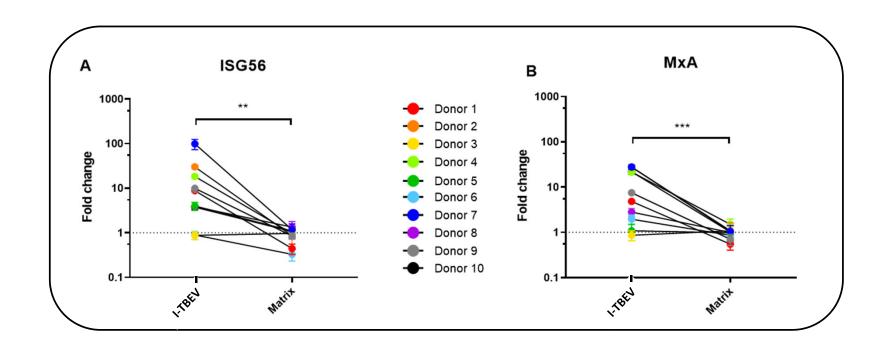


- Linear relationship between % of conforming product and response
- 20% difference results in statistically significant differences in response





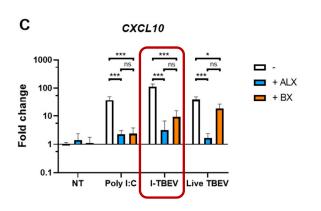
PBMCs from different donors respond in a qualitatively similar manner (though quantitatively different)



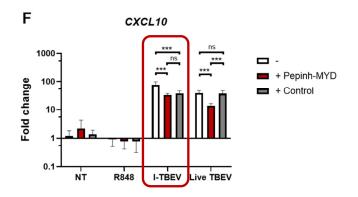




Activation of cryopreserved PBMCs relies on triggering of the RIG-I pathway



Inhibitors interfering with RIG-I pathway block I-TBEV induced responses



Inhibitors of TLR signaling have no specific effects





Conclusions

- In vitro systems mimicking diverse aspects of the immune system are available
- Simple PBMC-based systems allow the study of vaccine-induced effects on DCs, conventional CD4 and CD8 T cells, and T follicular helper cells; B cells are currently under investigation
- DCs and T cells show distinct responses to different types of vaccines which correlate with responses in vivo
- PBMCs can detect differences in vaccine quality with high resolution and can be used in the context of batch quality control





ACKNOWLEDGEMENTS

- Aurora Signorazzi
- Gabriela Tapia Calle
- Shuran Gong
- Jacqueline de Vries-Idema
- Anke Huckriede

University Medical Center Groningen Groningen, The Netherlands









umcg







Istituto Superiore de Sanitá, Rome, Italy

Jeroen Pennings



Nat. Inst. Public Health and the Environment Bilthoven, The Netherlands



