



Latvian Biomedical  
Research and Study Centre  
research and education in biomedicine from genes to human

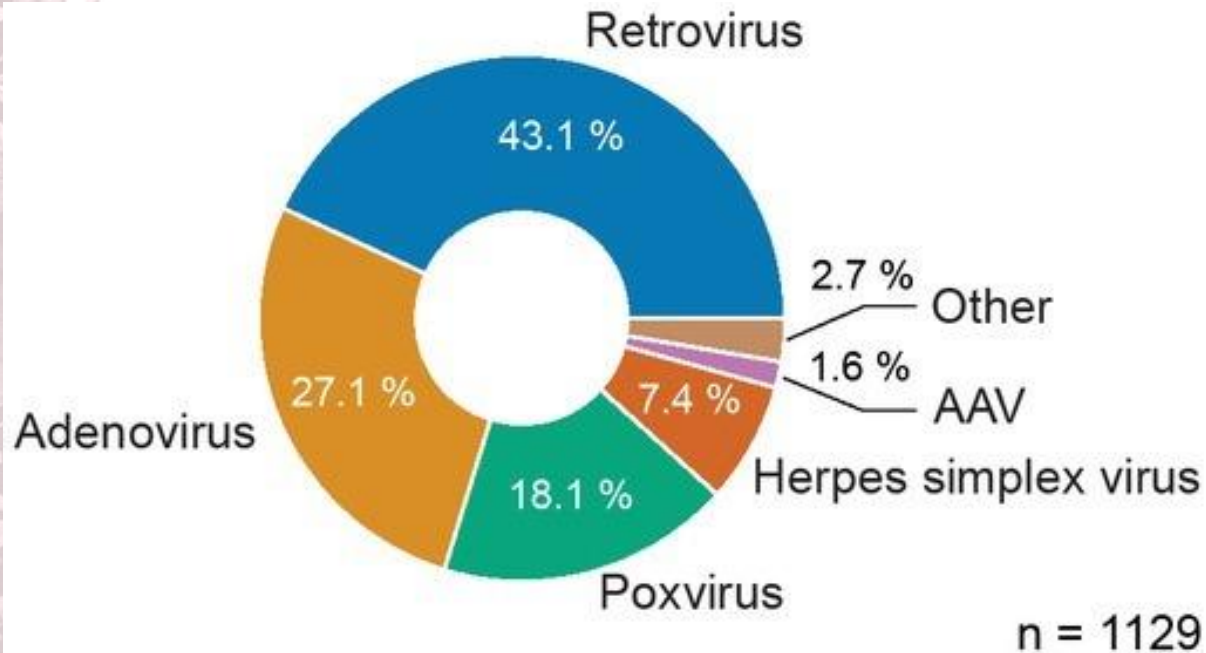
# THERAPEUTIC MODULATION OF TUMOR MICROENVIRONMENT WITH RECOMBINANT VIRAL VECTORS

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# Application of viral vectors in clinical trials to treat cancer

## % of open clinical trials



Bezeljak, 2022

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8884858/>

Pier Francesco Ferrucci, Laura Pala, Fabio Conforti, and Emilia Cocorocchio. Talimogene Laherparepvec (**T-VEC**): An Intralesional Cancer Immunotherapy for Advanced Melanoma. *Cancers (Basel)*. 2021 Mar; 13(6): 1383. 2021 Mar 18. doi: 10.3390/cancers13061383

Zhang W-W, Li L, Li D, Liu J, Li X, Li W, et al. The first approved gene therapy product for cancer **Ad-p53 (Gendicine)**: 12 years in the clinic. *Human Gene Therapy*. 2018;29:160-179. DOI: 10.1089/hum.2017.218

Liang M. **Oncorine**, the world first oncolytic virus medicine and its update in China. *Current Cancer Drug Targets*. 2018;18:171-176. DOI: 10.2174/1568009618666171129221503

## Delytact

Jahan N, Ghouse SM, Martuza RL, Rabkin SD. In situ cancer vaccination and immunovirotherapy using oncolytic **HSV**. *Viruses*. 2021;13:1–27. doi: 10.3390/v13091740.

## phase III clinical study

IFN $\alpha$ -2b rAd-IFN $\alpha$ /Syn3 (Ad5)+ polyamid surfactant (Syn3); patients with non-muscle-invasive bladder cancer (Boorjian, et al. Intravesical Nadofaragene Firadenovec Gene Therapy for BCG-Unresponsive Non-Muscle-Invasive Bladder Cancer: A Single-Arm, Open-Label, Repeat-Dose Clinical Trial. *Lancet Oncol*. 2021, 22, 107–117.)

**GM-CSF T-VEC** ( $\Delta$ ICP34.5  $\alpha$ HSV1) **anti-CTLA-4** (ipilimumab) clinical trials with metastatic stage IIIB/C–IVM1a melanoma (Ferrucci, et al, Talimogene Laherparepvec (T-VEC): An Intralesional Cancer Immunotherapy for Advanced Melanoma. *Cancers* 2021, 13, 1383.

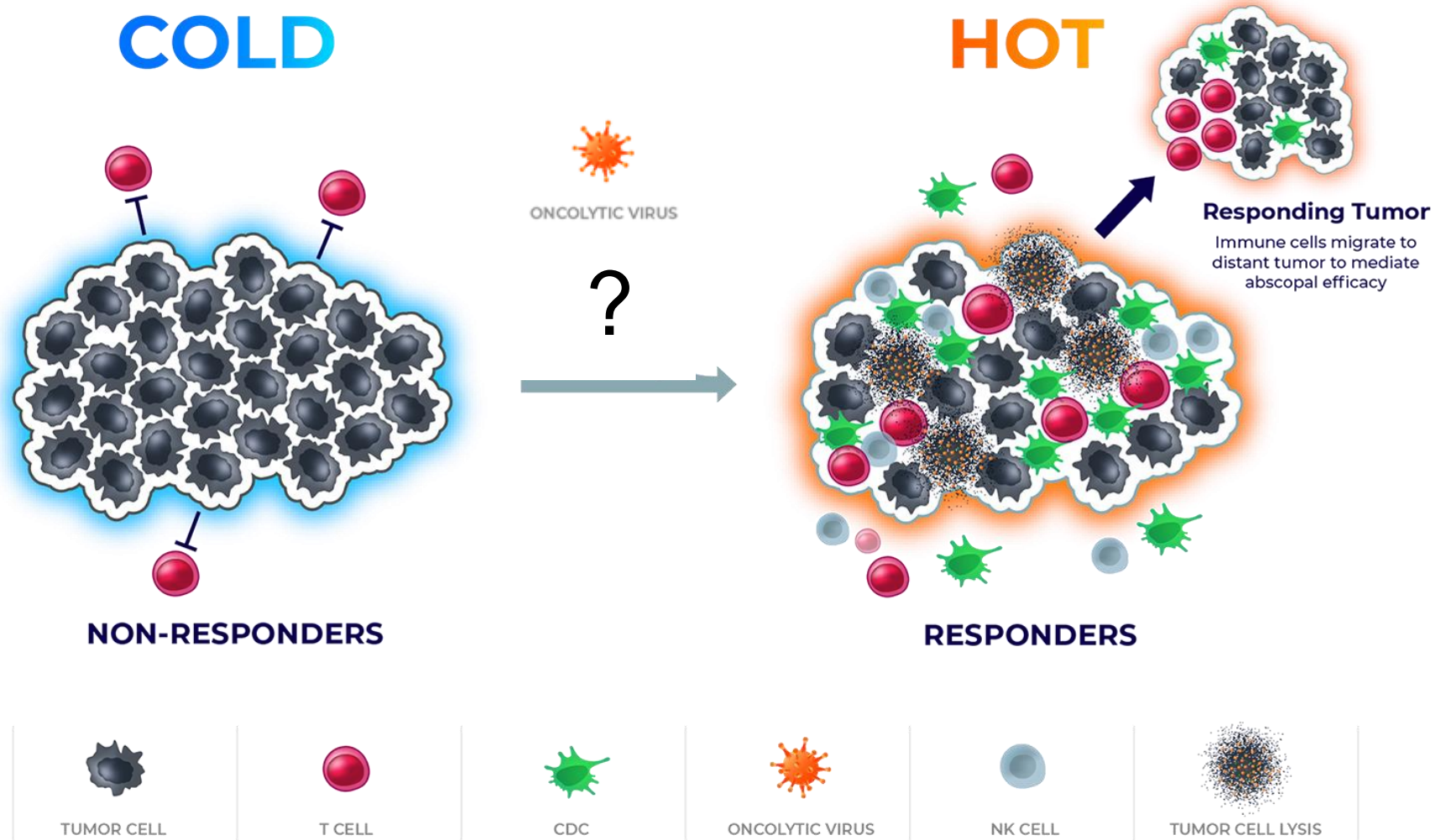


Table 1. Comparison of viral vectors.

Virus Type	Insert Capacity	Cell Receptor/Tropism	Advantages	Limitations
DNA vectors				
Adenoviruses	up to 7.5 kb up to 36 kb (fully deleted helper dependent Ads)	Coxsackie Adenovirus receptor (CAR); CD46	<ul style="list-style-type: none"> <li>• high transduction efficiency</li> <li>• broad tissue tropism</li> <li>• availability of scalable production system</li> <li>• tumour-specific gene promoters</li> </ul>	<ul style="list-style-type: none"> <li>• pre-existing viral immunity</li> <li>• strong immune responses against vector proteins</li> <li>• biosafety concerns (random integration)</li> </ul>
Poxviruses	up to 24 kb 7.5 kb (MVA)	binding to glycosaminoglycans following cell fusion; virus replication and spread are dependent on epidermal growth factor receptor (EGFR) signalling; preferential replication in cancer cells	<ul style="list-style-type: none"> <li>• inherently tumour targeting</li> <li>• cytoplasmic replication</li> <li>• low prevalence of anti-vector immunity</li> <li>• large production of clinical grade preparations available</li> <li>• stable in blood following intravenous injection and highly efficient systemic delivery</li> </ul>	<ul style="list-style-type: none"> <li>• replication-deficient Poxvirus vectors encoding heterologous antigens have a lower ability to prime immune responses in humans than other viral vectors</li> <li>• adaptive immune response against the vector</li> <li>• large virus particles hampering their intratumoral spread</li> </ul>
Herpesviruses	up to 40 kb (replication-deficient vector) up to 14 kb (HSV1)	Herpesvirus Entry Mediator and nectin 1 (HSV-1)	<ul style="list-style-type: none"> <li>• selective replication in tumours</li> <li>• potent cytolytic capability</li> <li>• blood-brain barrier crossing</li> <li>• availability of scalable production system</li> </ul>	<ul style="list-style-type: none"> <li>• potential neurovirulence (HSV-related encephalitis)</li> <li>• genetically modified HSV vectors are not very efficient compared to oncolytic wild-type variants</li> <li>• pre-existing immune response</li> <li>• strong immune responses against vector proteins</li> </ul>

Spunde et al,  
*Biomedicines*, 2022



Virus Type	Insert Capacity	Cell Receptor/Tropism	Advantages	Limitations
RNA vectors				
Rhabdoviruses	4–6 kb	multiple receptors were proposed (phospholipids and gangliosides, nicotinic acetylcholine receptor, neural cell adhesion molecule, and low-density lipoprotein gene family receptors (LDLR))	<ul style="list-style-type: none"> <li>selective and efficient replication in tumour cells including in metastases</li> <li>high oncolytic properties</li> <li>pseudotyping capabilities</li> <li>ability to cross the blood-brain barrier (VSV)</li> </ul>	<ul style="list-style-type: none"> <li>infection of tumour-associated DCs reduces their antigen presentation properties</li> <li>potential neurovirulence (VSV)</li> <li>insufficiently developed large-scale manufacturing technology</li> </ul>
Alphaviruses	up to 5 kb	very low-density lipoprotein receptor (VLDL-R) and apolipoprotein E receptor 2 (ApoER2)	<ul style="list-style-type: none"> <li>low specific immune response against the vector</li> <li>low pre-immunity</li> <li>tumour tropism (SIN)</li> <li>induction of immunogenic cell death</li> <li>high level of transgene expression</li> </ul>	<ul style="list-style-type: none"> <li>modest insert capacity</li> <li>short time expression potential</li> <li>neurovirulence (SFV)</li> <li>insufficiently developed large-scale vector production system</li> </ul>
Arenaviruses	up to 2 kb	preferentially infect monocytes, macrophages, and DCs through binding to $\alpha$ -dystroglycan ( $\alpha$ -DG)	<ul style="list-style-type: none"> <li>non-lytic infection of dendritic cells</li> <li>efficient CD8<sup>+</sup> T cell immunity</li> <li>weak neutralizing antibody response against the vector</li> <li>rare pre-existing anti-vector immunity</li> <li>safe in human</li> </ul>	<ul style="list-style-type: none"> <li>low insert capacity</li> <li>limited direct oncolytic properties</li> <li>insufficiently developed large-scale vector production system</li> </ul>

Virus Type	Insert Capacity	Cell Receptor/Tropism	Advantages	Limitations
RNA vectors				
Enteroviruses	0.3–1.7 kb	immunoglobulin-like receptor, CD155; Nectin-like molecule 5 (Poliovirus), coxsackie-adenovirus receptor (CAR); RGD motif of integrins (Coxsackievirus), other co-receptors	<ul style="list-style-type: none"> <li>tumour tropism</li> <li>neurotropism (Poliovirus)</li> <li>low pathogenicity</li> <li>oncolytic replication</li> </ul>	<ul style="list-style-type: none"> <li>very low insert capacity</li> <li>unstable genome</li> <li>high levels of pre-existing immunity to polio vectors</li> <li>insufficiently developed large-scale vector production system</li> </ul>
Reoviruses	up to 1.5 kb within two RNA segments	the receptor is unknown, but is thought to include sialic acid and junctional adhesion molecules (JAMs)	<ul style="list-style-type: none"> <li>oncolytic properties</li> <li>relatively non-pathogenic in adults</li> <li>tumour tropism</li> </ul>	<ul style="list-style-type: none"> <li>insufficiently developed recombinant vector platform</li> <li>transient expression</li> <li>anti-vector pre-immunity (neutralising Abs)</li> <li>very low insert capacity</li> </ul>
Paramyxoviruse	up to 6 kb (Measles virus, MV) 4.5 kb (Newcastle disease virus, NDV)	different receptors: MV: signal lymphocyte-activation molecule (SLAM or CD150) CD46, Nectin-4 NDV: sialic acids on the tumour cell surface	<ul style="list-style-type: none"> <li>tumour tropism</li> <li>pDC maturation</li> <li>oncolytic properties</li> <li>low seroprevalence (NDV)</li> <li>safe for human</li> </ul>	<ul style="list-style-type: none"> <li>pre-existing immunity (MV)</li> <li>insufficiently developed recombinant vector platform</li> </ul>

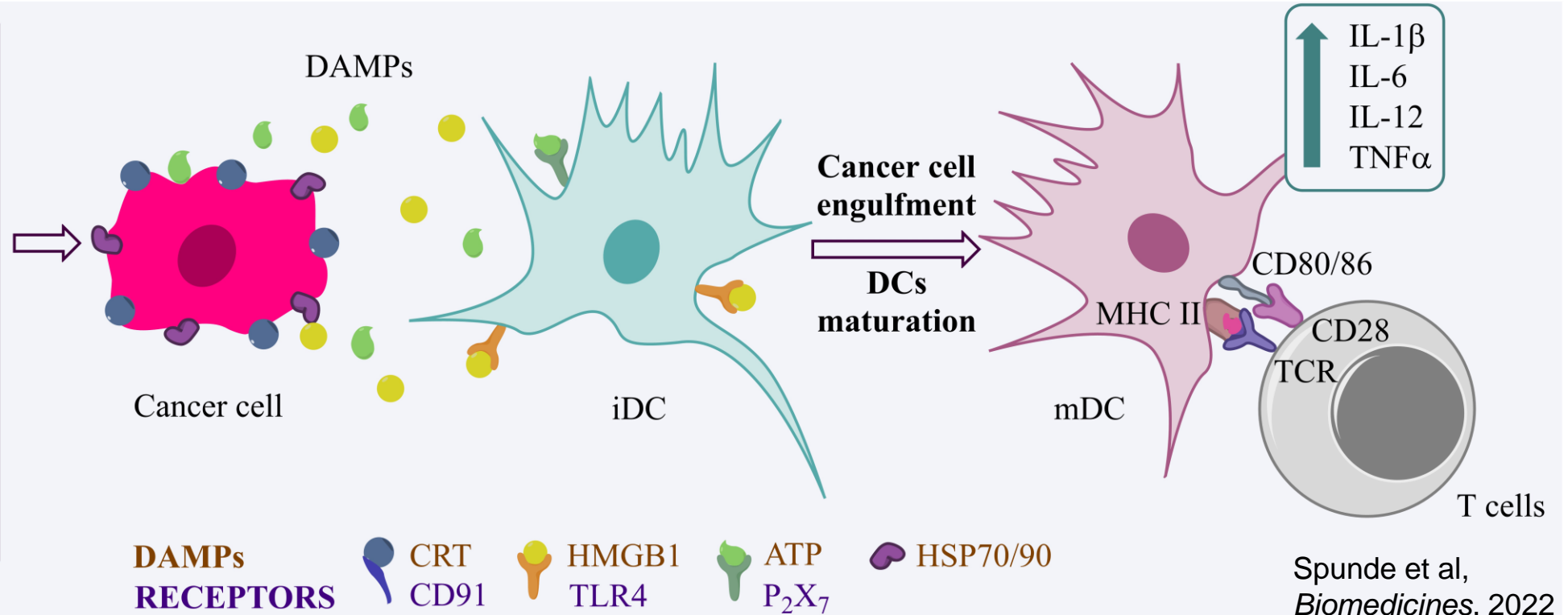
## Immunogenic cell death (ICD)

### VIRAL INFECTION

*Measles virus,  
Coxsackievirus B3,  
Semliki Forest virus,  
Newcastle disease virus*

### PHYSICAL STRESS

### CHEMOTHERAPY



Ma et al. *Cell Death and Disease* (2020)11:48  
<https://doi.org/10.1038/s41419-020-2236-3>

Cell Death & Disease

### ARTICLE

### Open Access

## Characterization of virus-mediated immunogenic cancer cell death and the consequences for oncolytic virus-based immunotherapy of cancer

Jing Ma<sup>1</sup>, Mohanraj Ramachandran<sup>1</sup>, Chuan Jin<sup>1</sup>, Clara Quijano-Rubio<sup>1,2</sup>, Miika Martikainen<sup>1</sup>, Di Yu<sup>1</sup> and Magnus Essand<sup>1</sup>

**Adenovirus** initiates multiple cell death pathways including necroptosis, inflammasome activation and autophagy before the tumor cells die by Ad-mediated lysis.

**VV** mediated cell lysis is primarily accompanied by induction of necroptosis and autophagy

**SFV4** induces rapid cell lysis accompanied by induction of immunogenic apoptosis

Necroptosis is not associated with SFV4 infection



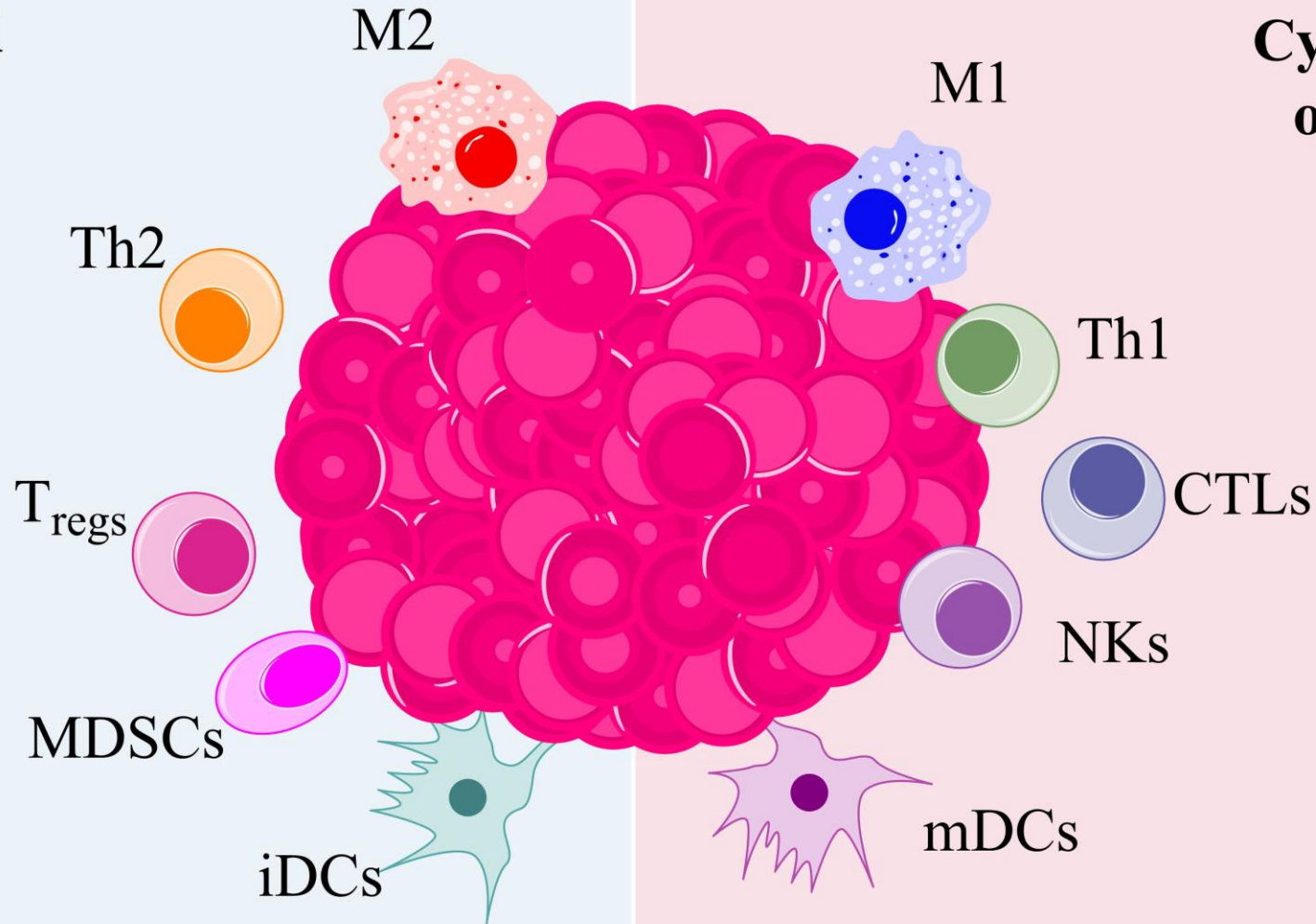
# Tumour Immune Microenvironment

**"Cold"**  
**Immunosuppressive**

**"Hot"**  
**Immunostimulating**

**Cytokines and  
other factors**

IL-4  
IL-10  
IDO  
COX2  
EGF  
HGF  
TGF $\beta$   
VEGF



**Cytokines and  
other factors**

NO  
IL-1 $\beta$   
IL-6  
IL-12  
TNF $\alpha$   
IFN $\gamma$



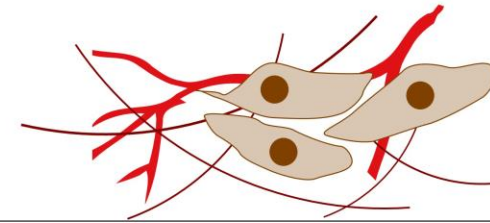
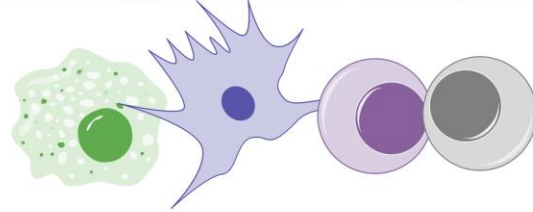
# Therapeutic strategies

①

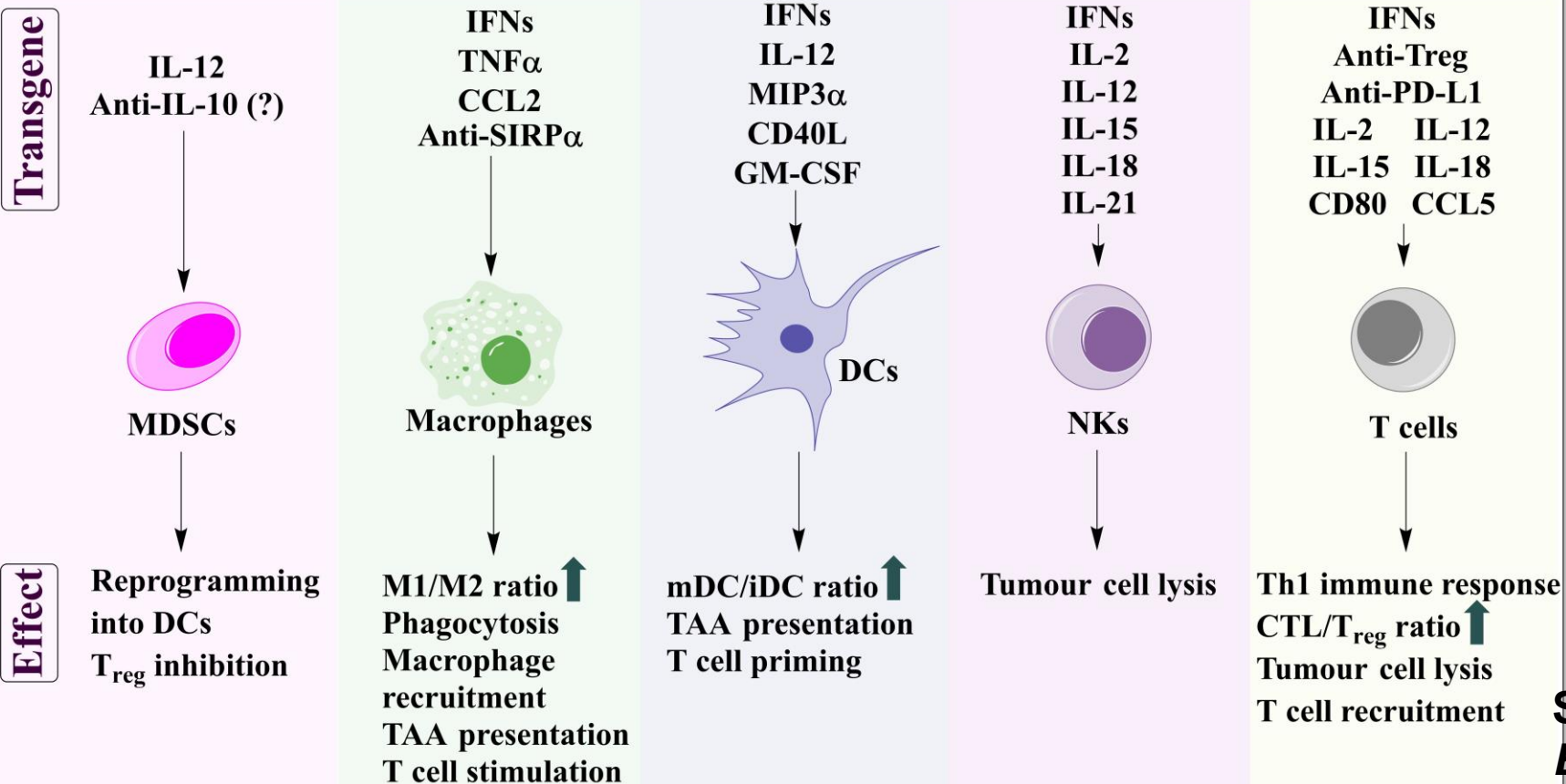
②

## Programming of tumour immune cells

## Programming of tumour stroma



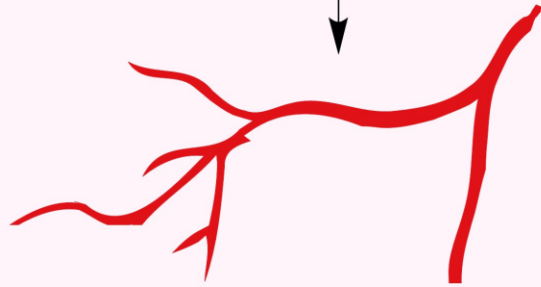
## Programming of tumour immune cells



## Programming of tumour stroma

Transgene

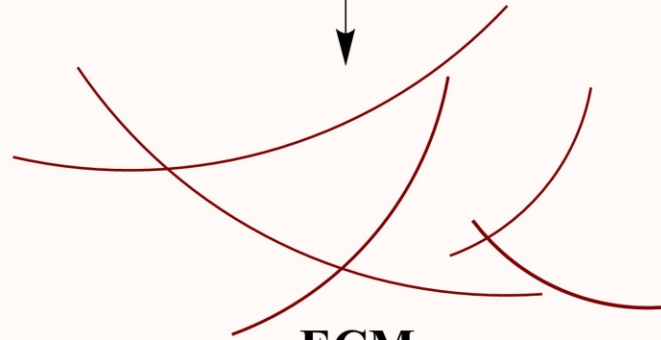
Anti-TGF $\beta$   
Anti-VEGF



Vascular endothelial cells

Angiogenesis ↓

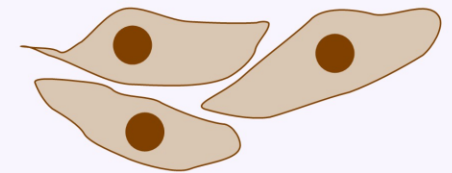
Relaxin  
Hyaluronidase



ECM

Intratumoral virus spread,  
drug delivery and immune  
cell infiltration ↑

Anti-TGF $\beta$



CAFs

Invasion ↓

Effect

2

Table 4. Virus vectors encoding cytokines and other molecules for remodelling tumour multiple immune cell populations.

Transgene	Virus Vector (Virus Backbone)	Model	Effect
Adenoviruses			
TNF $\alpha$ IL-2	Ad5-CMV- mTNF $\alpha$ Ad5-CMVmIL2 (Ad5)	murine B16.OVA melanoma; C57 BL/6JOLA Hsd mice	complete tumour regression in all animals treated with anti-PD1 antibodies and corresponding viruses; Th1 immune response and increased intra-tumoral proportion of CD8 <sup>+</sup> and CD4 <sup>+</sup> T cells
GM-CSF CTLA-4 ab	SKL001 SKL002 Ad5	CMT-64 mouse small lung carcinoma, B16F10 murine melanoma, human A549 lung s.c. xenograft model	selective replication and anti-tumour activity after intravenous administration was shown in mouse B16F10 melanoma tumour and human tumour xenograft model; combination of the viruses potentiated anti-tumour activity
Anti TGF $\beta$ shRNA GM-CSF (in one vector) plus MART1 (DNA/TAA)	AdGshT	murine B16BL6-CAR/E1B55 malignant melanoma; C57BL/6 mice	treatment by both DNA vaccine expressing TAA (MART1) and oncolytic adenovirus, encoding GM-CSF together with shRNA to TGF- $\beta$ 2 resulted in significant anti-tumour effects, however, complete regression of tumours was not achieved
IL-12p35, IL-12p40; GM-CSF and RLX (relaxin)	oAd/RLX oAd/IL12/GM- RLX	Syrian hamster s.c. and orthotopic pancreatic tumour models.	expression of IL-12, GM-CSF and RLX mediated by a single oncolytic Ad vector promoted remodelling of TME to potentiate antibodies-based therapies
IL-12 plus VEGF binding shRNA	RdB/IL12/shVEGF (Ad5)	murine B16-F10 melanoma; C57BL/6 mice	Efficient anti-tumour effect with massive tumour infiltration of differentiated CD4 <sup>+</sup> T cells, CD8 <sup>+</sup> T cells, NK cells, and DCs. Suppressed expression of VEGF, supporting the restoration of the anti-tumour immune response



Herpesviruses			
CCL2 mIL-12	M010 M002 ( $\Delta$ ICP34.5 oHSV)	neuroblastoma Neuro-2a tumours s.c. syngeneic A/J mouse strain	combined treatment led to the most efficient tumour growth inhibition
IL-12 IL-15 PD1v GM-CSF IL7 plus CCL19	oHSV2-IL12, -PD1v, -IL15, -IL7-CCL19, -GM-CSF $\Delta$ ICP34.5 $\Delta$ ICP47 oHSV2 (HG52 strain)	breast cancer 4T1 and colon carcinoma CT26 murine tumour models; Balb/c mice	all vector variants used as a single treatment have had a similar anti-tumour activity; the most potent activity was demonstrated for all five virus vector combinations; the tumour re-challenge exhibited that cocktail therapy prevents secondary tumourogenesis
IL-12 GM-CSF (in one vector)	R-123 hHER2 retargeted $\Delta$ ICP34.5 oHSV	human breast cancer SK-OV-3 cells, Lewis lung carcinoma murine cell line expressing hHER2 (HER2-LLC1) s.c. tumours; hHER2-transgenic C57BL/6 mice (B6.Cg-Pds5bTg(Wap ERBB2)229Wzw/J)	combined treatment with anti-PD1 led to significant inhibition of tumour growth with complete tumour resection in case, (mGM-CSF), mIL-12+mGM-CSF) expressing vector; systemic delivery of double-armed virus combined with anti-PD1 inhibited the development of tumour metastasis
Poxviruses			
PD-1 fused with IgG1 Fc plus GM-CSF	VV-iPDL1/GM WR vvDD	murine s.c. tumours Luc B16-F10 melanoma; Murine breast cancer Py230 and MC38 colon adenocarcinoma; C57BL/6 mice	the highest tumour growth inhibition was observed in VV-iPDL1/GM treated animals, compared to single treatments; CD8 T cell depletion significantly abolished the systemic anti-tumour activity of VV-iPDL1/GM; increased DCs (CD11c <sup>+</sup> ) infiltration was observed in VV-iPDL1/GM treated mice

# Alphaviruses

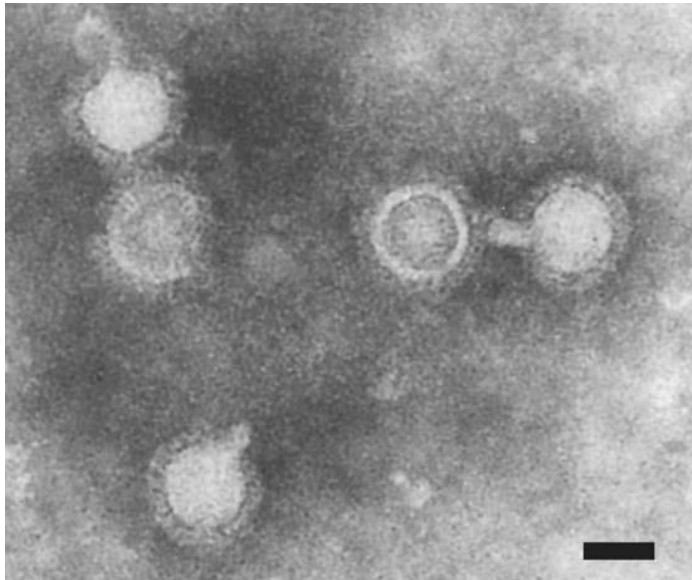
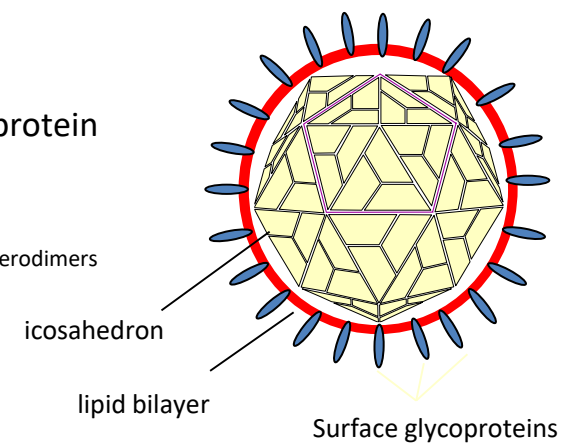
Enveloped virus

Icosahedron : 240 copies of 1 protein

Spherical : 65-70nm

Envelope : 80 trimer spikes

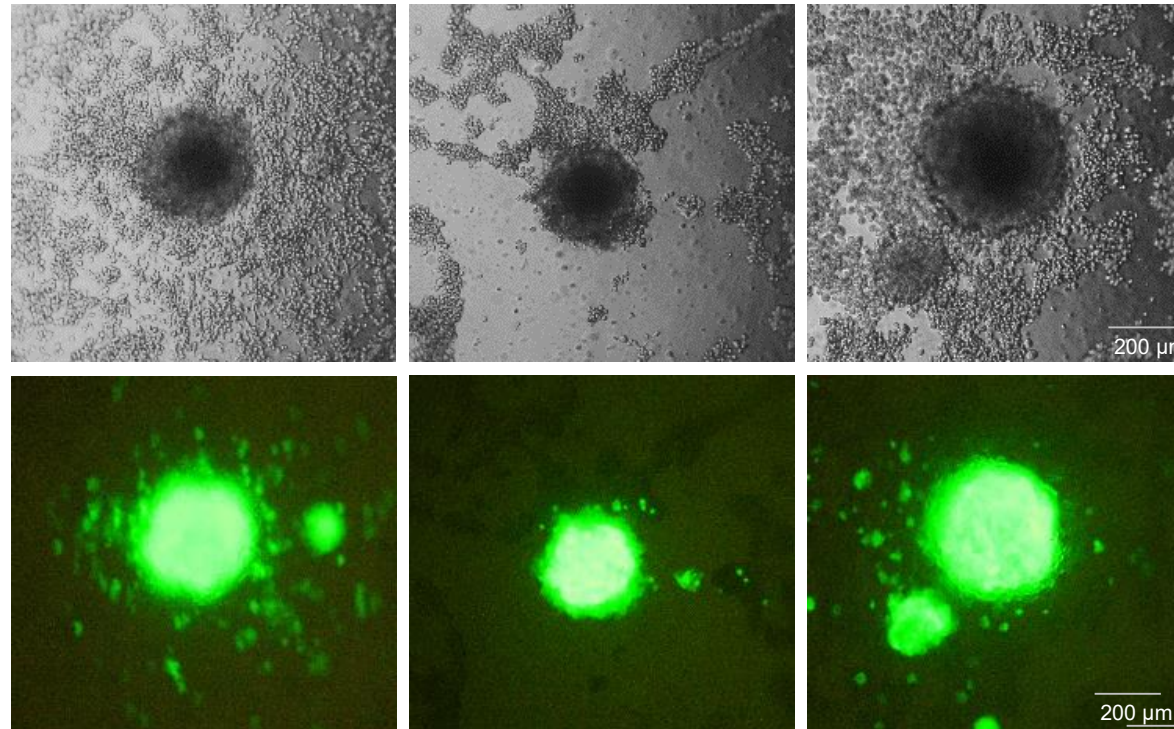
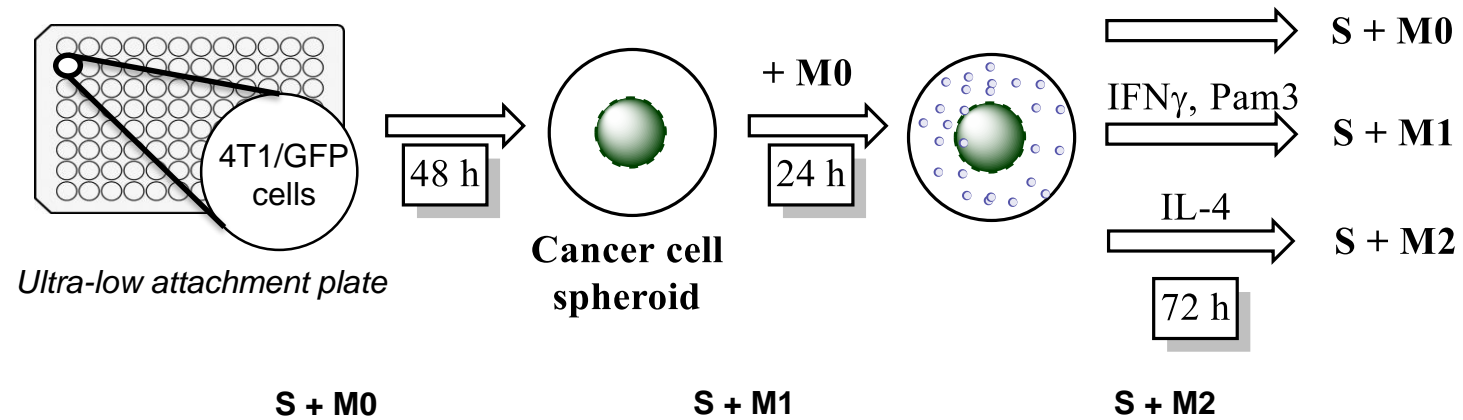
each spike = 3 x E1/E2 heterodimers



Electron microscopy of recombinant SFV particles. Negative staining, bar 50 nm.  
(Zajakina *et al*, 2010)

- RNA replication and high transgene expression
- No risk of integration
- Alphaviruses can target lymph nodes
- Dendritic cells infection
- Targeting cancer cells
- High virus titers
- Safe for human
- No vector preimmunity
- Transient expression
- Oncolytic properties
- Induce immunogenic cell death**

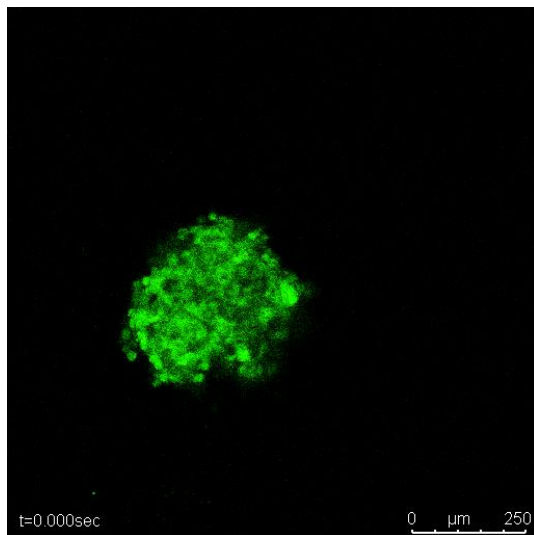
# Co-culture of 4T1/GFP cell spheroids with BMDM



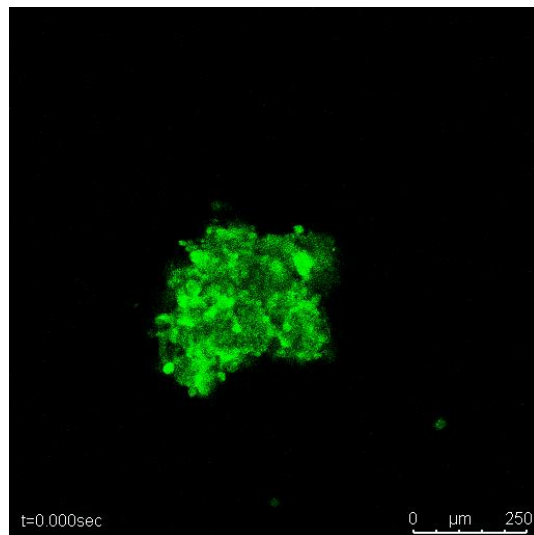


4T1/GFP (green), 48 h incubation with macrophages (unlabeled)

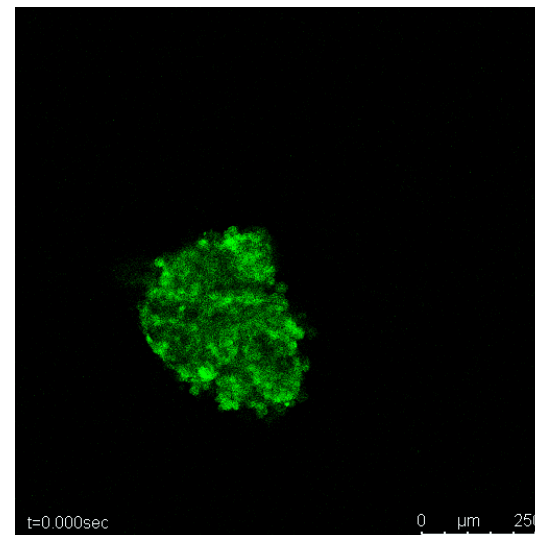
1\_6 control (w/o M)



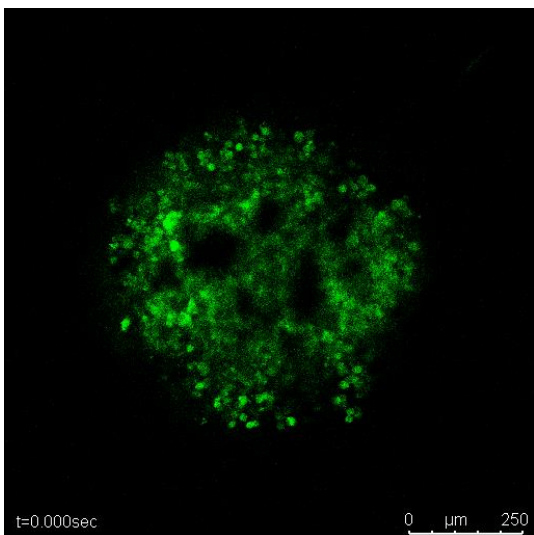
1\_9 M0



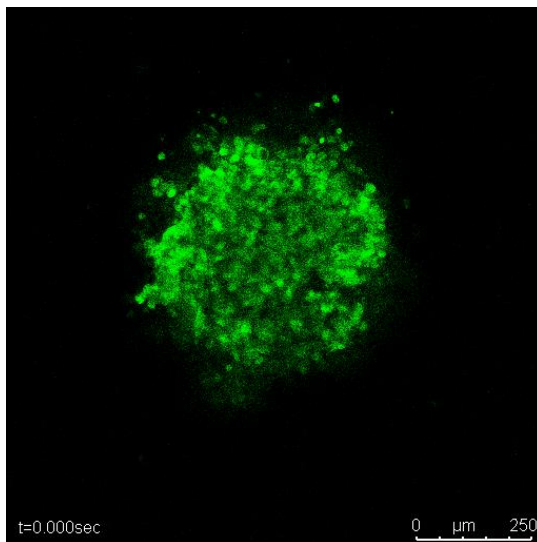
1\_4 M1



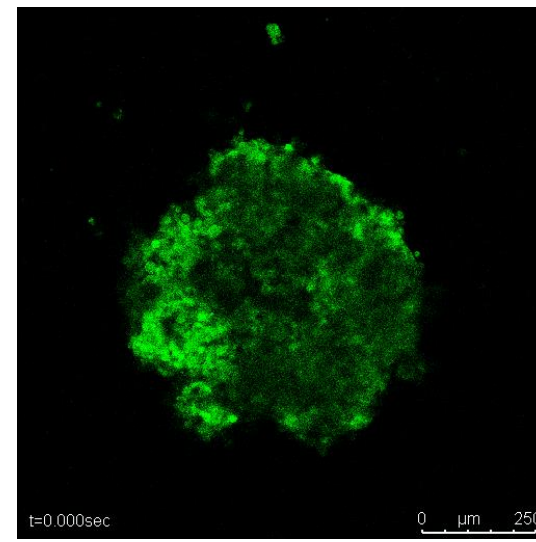
2\_2 control



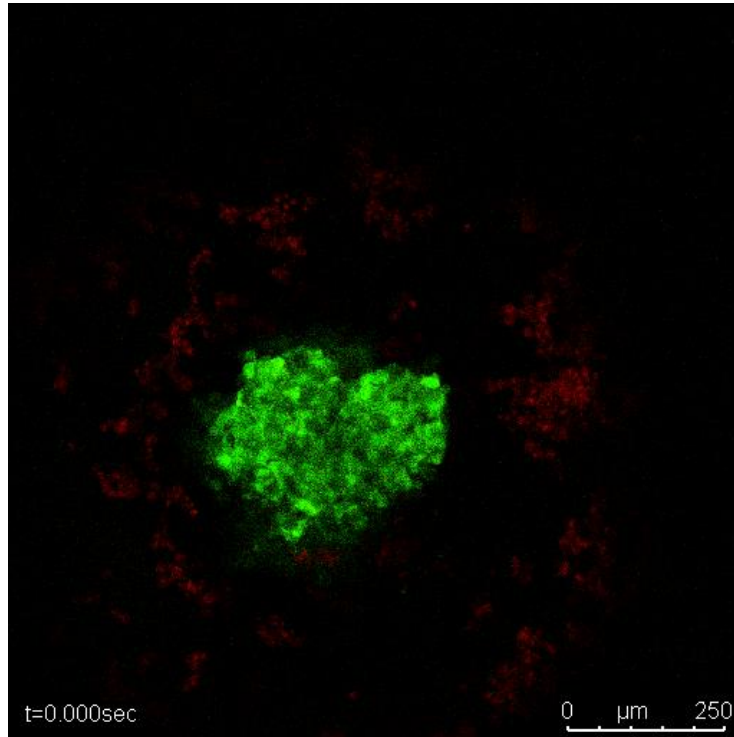
2\_9 M0



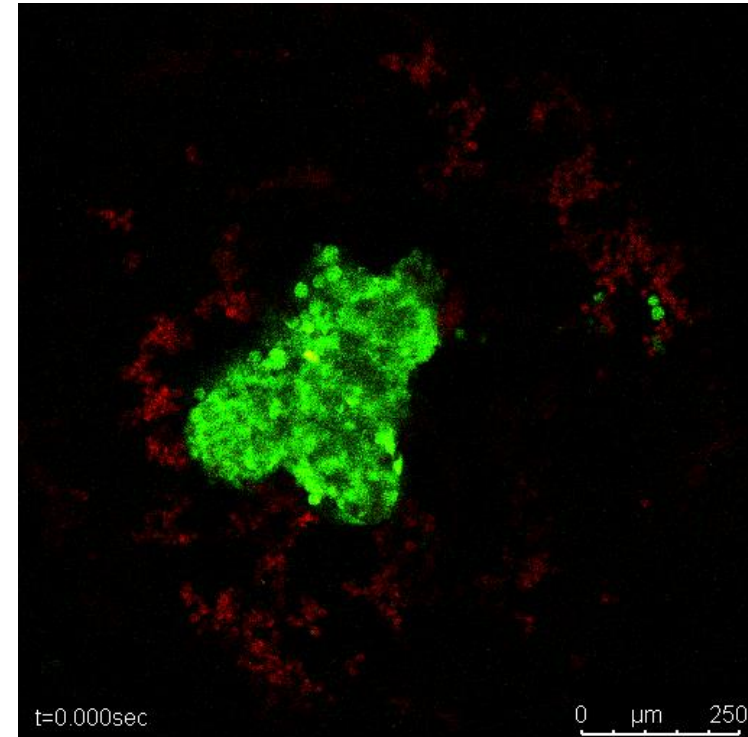
2\_4 M1



4T1/GFP (green) + M1 (red)



4T1/GFP (green) + M0 red

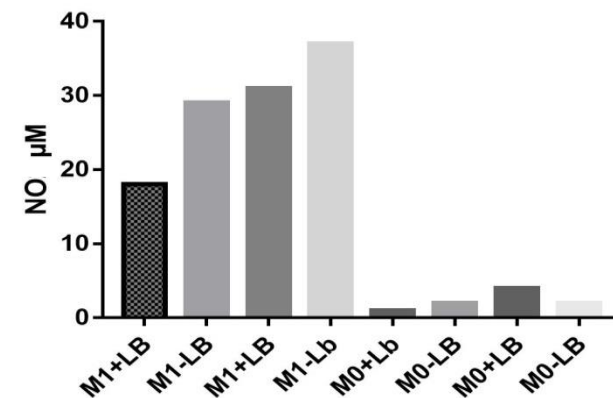


tumour spheroids (green) were incubated with macrophages (red) for 24 h

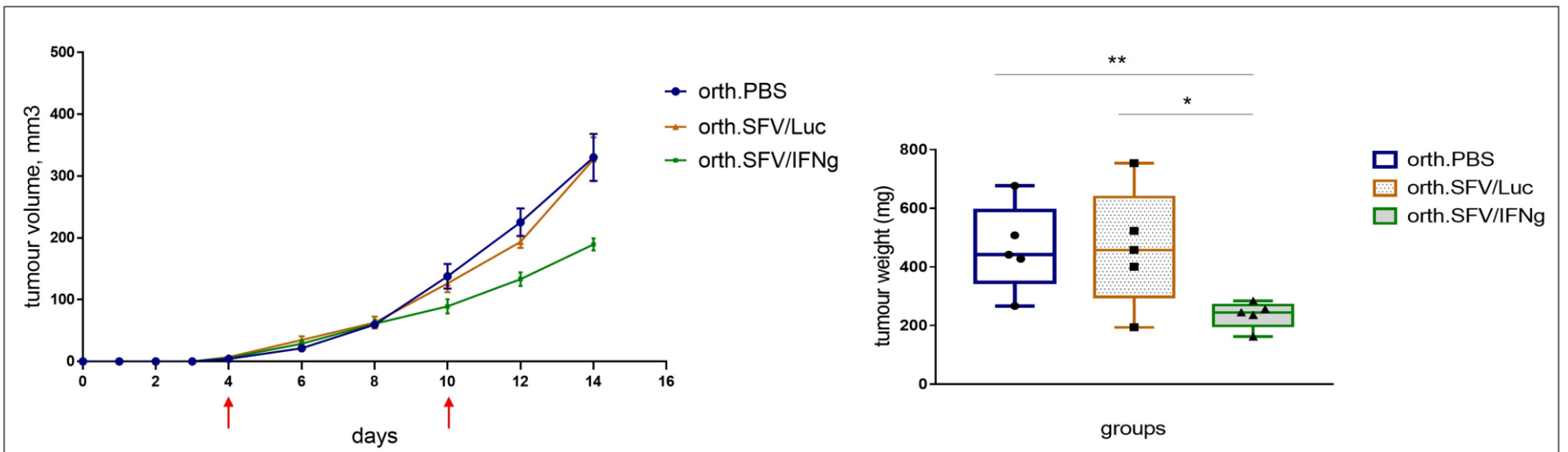
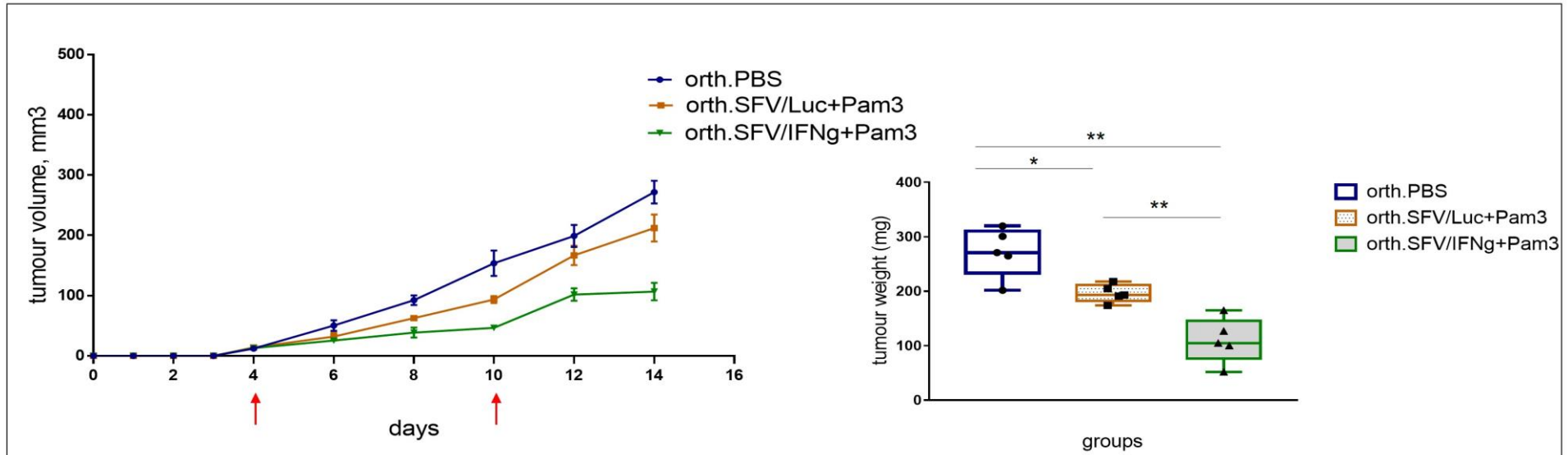
#### Observations:

- downregulation GFP intensity (M1)
- migration of GFP+ cells out of the spheroid (M0)
- NO test day 4

**NO test for sferoids**

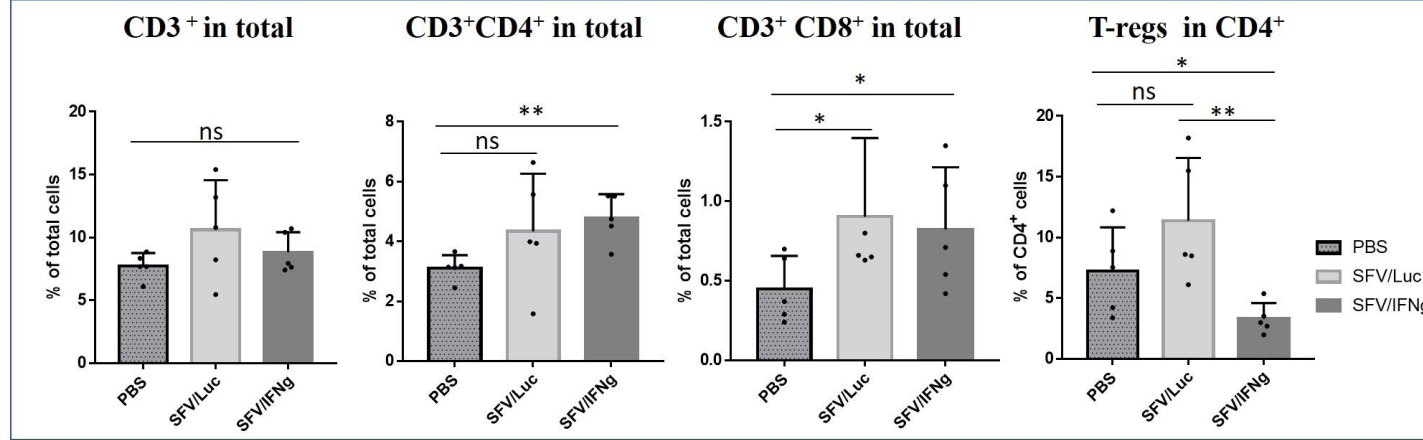


## Inhibition of 4T1 tumour growth by i.t. injection of SFV/IFN $\gamma$ virus

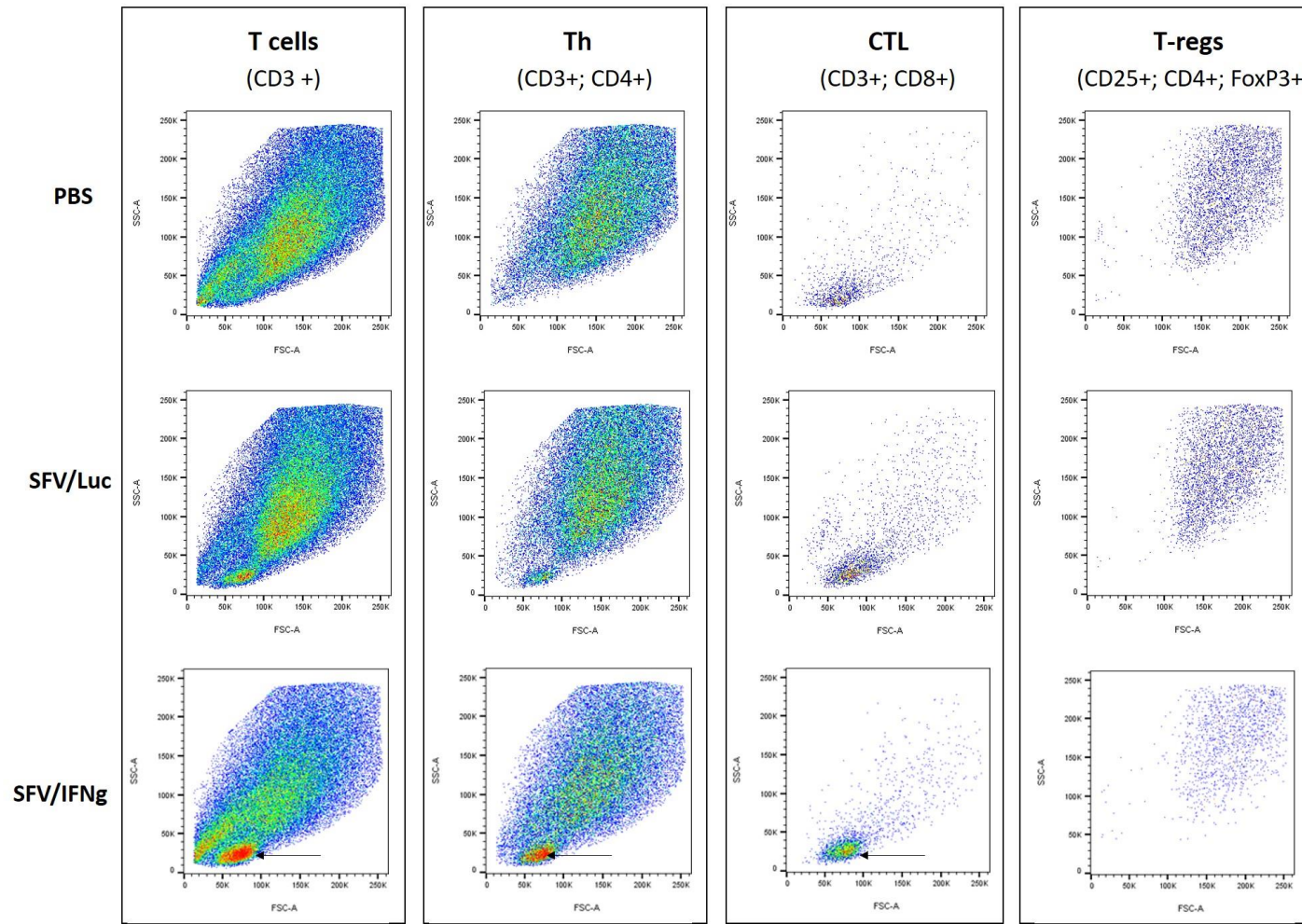




a.



b.

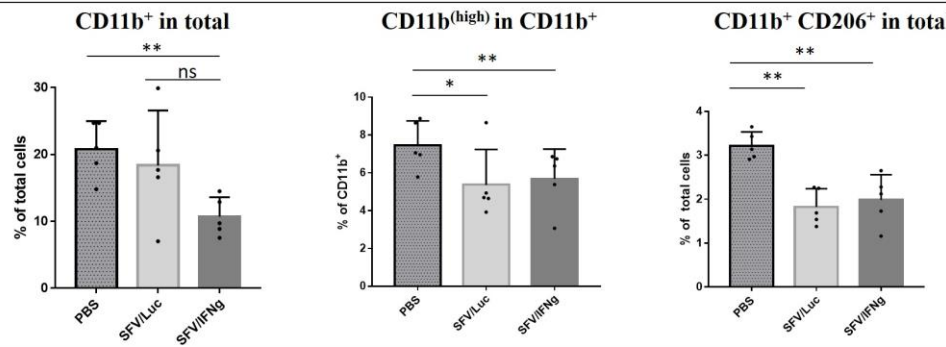


Flow cytometry analysis of T cells in the tumours treated with SFV/IFNg, SFV/Luc or PBS in orthotopic 4T1 mouse breast cancer model

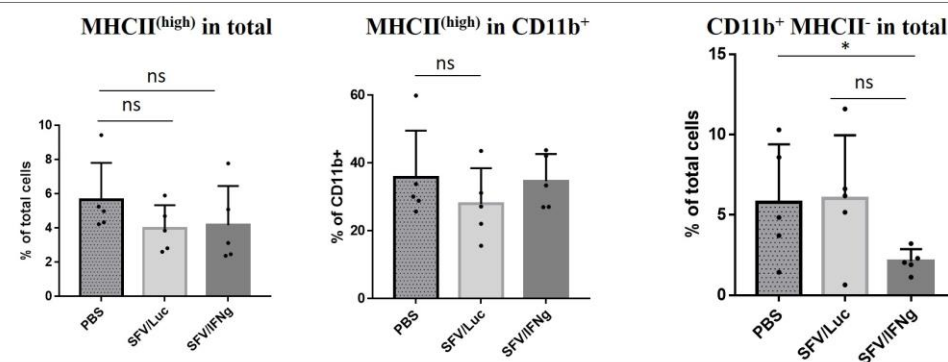


Flow cytometry analysis of myeloid cells in the tumours treated with SFV/IFN $\gamma$ , SFV/Luc or PBS in an orthotopic 4T1 mouse breast cancer model

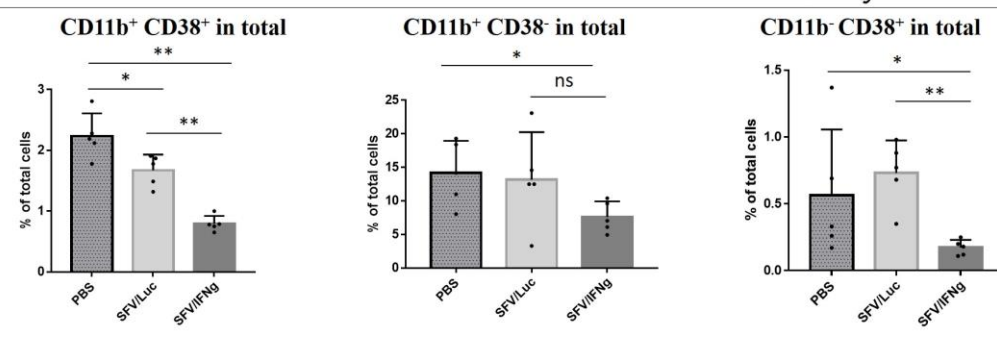
a.



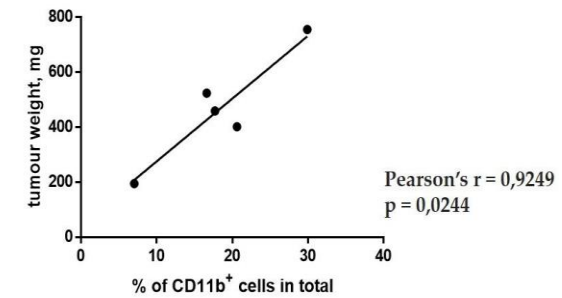
b.



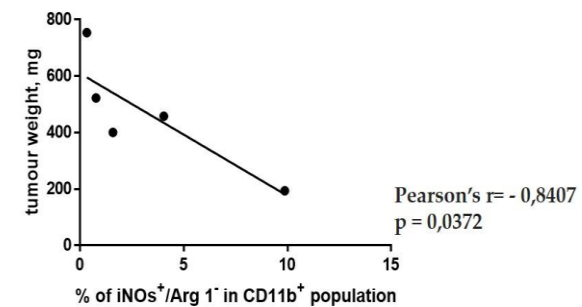
c.



correlation analysis (SFV/Luc group)

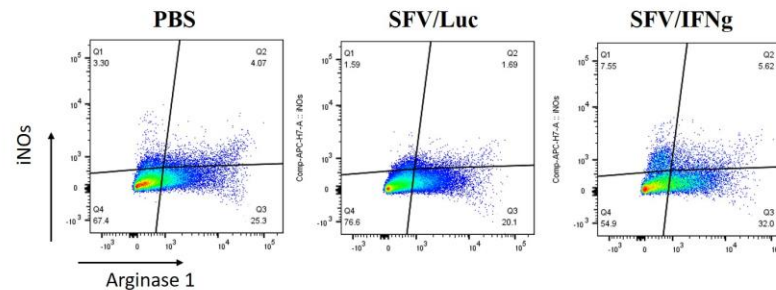
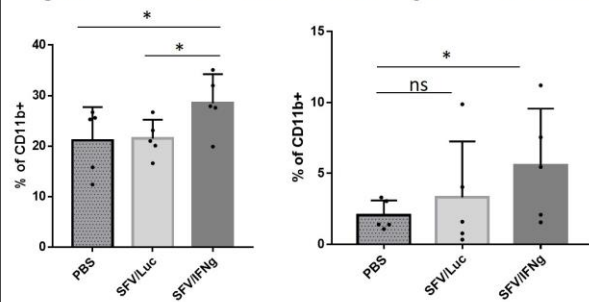


correlation analysis (SFV/Luc group)

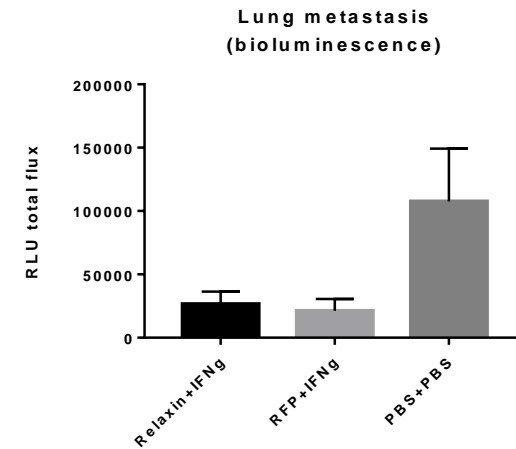
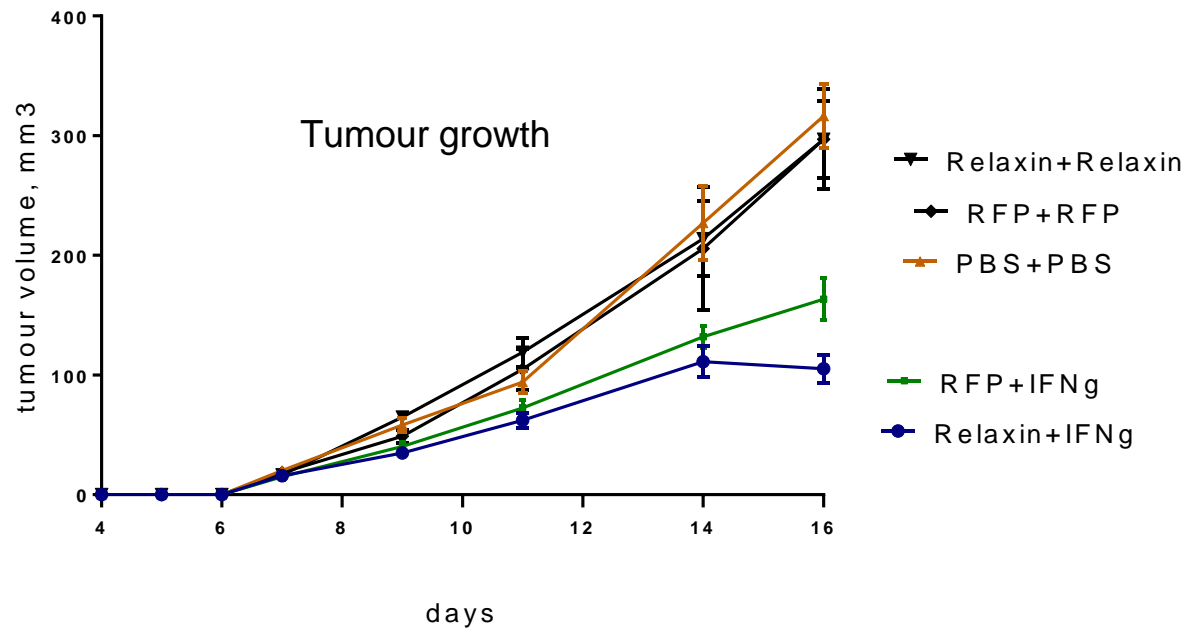
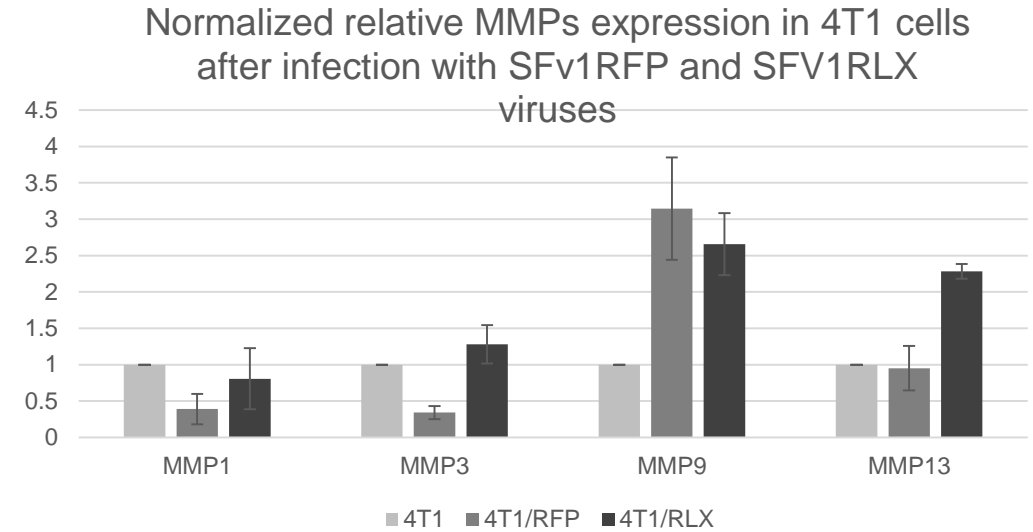
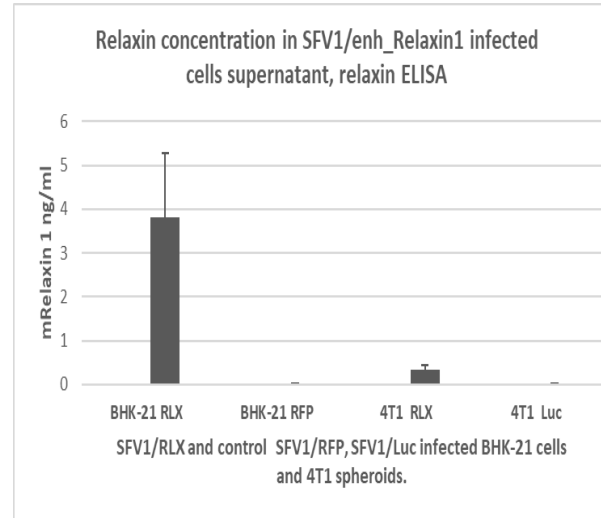
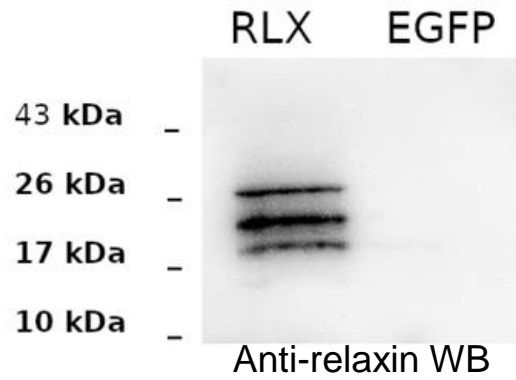


d.

Arginase 1<sup>+</sup> iNOs<sup>-</sup> in CD11b<sup>+</sup> iNOs<sup>+</sup> Arginase 1<sup>-</sup> in CD11b<sup>+</sup>



# Inhibition of 4T1 tumour growth by i.t. injection of SFV/IFN $\gamma$ virus in combination with SFV/Relaxin





- inhibition of tumor growth in an orthotopic 4T1 mouse breast cancer model
- significant increase in the populations of intratumoral Th cells and CTLs, and reduction of T-regs
- decreased intratumoral infiltration of myeloid cells expressing CD11b, CD206, or CD38
- enhanced inhibition of tumor growth in combination with Pam3CSK4 and Relaxin

SFV-based expression of IFN $\gamma$  benefits the antitumor immune response, representing a promising adjuvant to current immunotherapy and chemotherapy strategies.

## Acknowledgements

*Thank you!*

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## Group Members:

Olga Trofimova

Karina Spunde

Ksenija Korotkaja

Zhanna Rudeviča

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