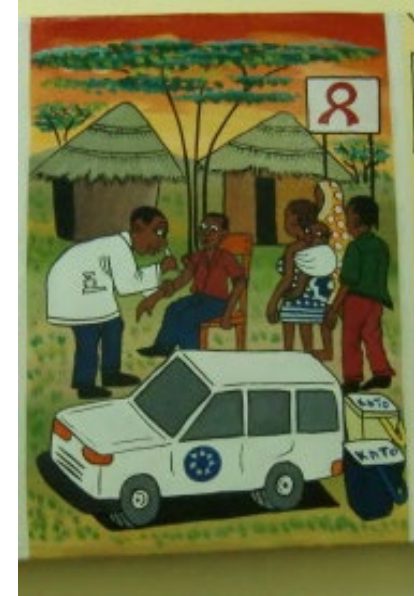


HIV vaccine-induced antibody responses impacts the accuracy of HIV testing algorithms in sub-Saharan Africa

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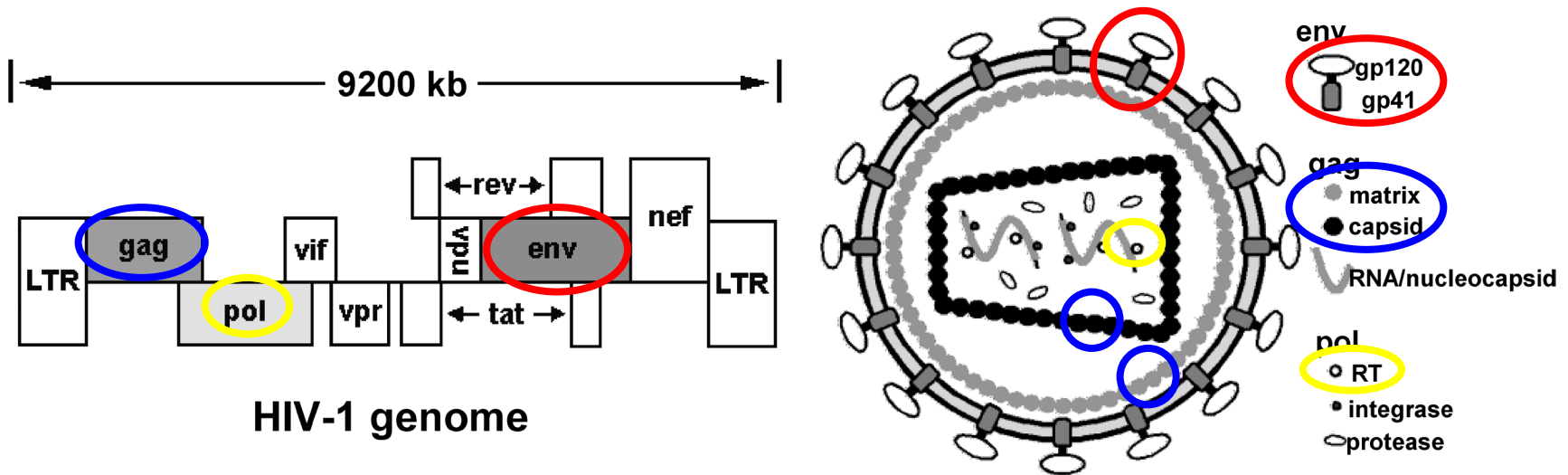
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Introduction

- A series of clinical HIV phase I/II vaccine trials were conducted in Sweden, Tanzania and Mozambique 2005 to 2015
- Prime-boost vaccine strategy using DNA-MVA-protein vaccines

Introduction

DNA-MVA-protein vaccine



HIV-1 genome

HIVIS-DNA



Env subtyp A,B,C

Gag subtyp A,B

Rtmut

rev B

HIVIS-DNA KI Wahren et al Mol Ther 2007;15:1724–1733



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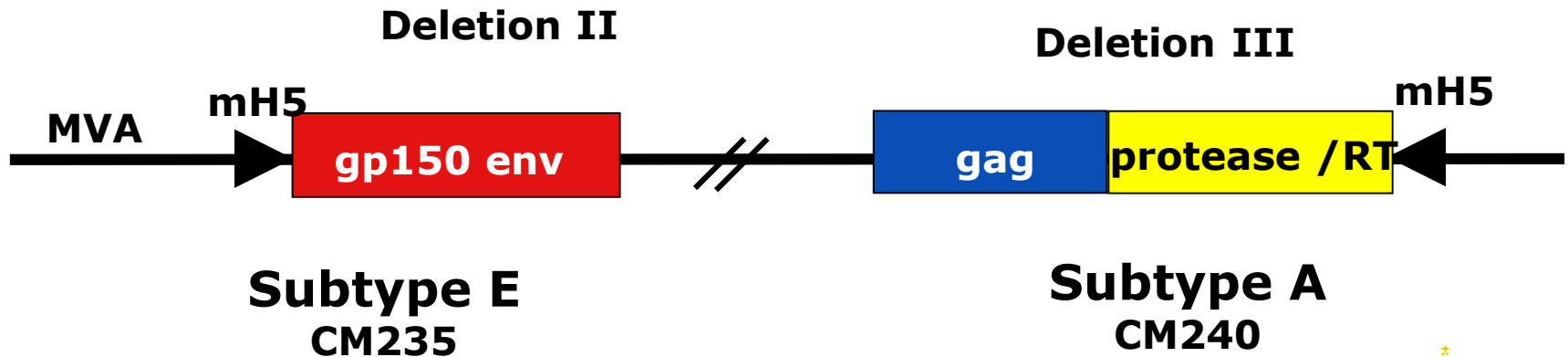
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Introduction

DNA-MVA-protein vaccine

Modified Vaccinia virus Ankara (MVA)/ Chiang Mai Double Recombinant (CMDR)

Developed by P Earl and B Moss, Laboratory of Viral Diseases, NIAID, NIH
Produced by Walter Reed Army Institute of Research



HIV-MVA-CMDR NIH/WRAIR Earl et al Vaccine 2009;27:5885–5895.



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Introduction

DNA-MVA-**protein** vaccine

- is a recombinant subtype C HIV-1 gp140 Env glycoprotein, CN54rgp140 adjuvanted with GLA-AF.
- GLA-AF is an adjuvant containing an aqueous formulation of glucopyranosyl lipid A, which is a synthetic monophosphoryl lipid A (MPL)-like molecule

Clegg CH, et al GLA-AF, an emulsion-free vaccine adjuvant for pandemic influenza. PLoS One. 2014; 9(2):e88979

Introduction

HIVIS01/02/05 phase I trial (3 HIV-DNA+2 HIV-MVA)

- All (100%) of 24 vaccinees developed antibodies to GAG
- All (100%) of 24 vaccinees were reactive in IMvHIV-1/HIV-2 III plus (Abbott) ELISA
- 13 (54%) of 24 were reactive in Enzygnost HIV Integral II ELISA
- 13 (54%) of 24 were also reactive in Western Blot (CDC criteria for positive classification; at least two bands of p24, gp41 or gp120/160)

Introduction

- Healthy uninfected HIV vaccine recipients will develop antibody responses that may result in diagnostic immunoassay reactivity, also known as vaccine-induced seroreactivity (VISR)
- HIV DNA or RNA PCR is used in all clinical HIV vaccine trials to rule out infection
- Volunteers in HIV vaccine trials carry a card identifying them as HIV vaccinees



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Folkhälsomyndigheten

Introduction

HIV diagnosis in resource-restricted countries depend on rapid diagnostic tests (RDTs)

Determine HIV1/2



15 min

Antigens: HIV-1 and HIV-2
Recombinant protein (RP)
and synthetic peptide

Antibodies: IgG

SD Bioline
HIV1/2 3.0



10-20 min

Recombinant
HIV-1 gp41, p24
HIV-2 p36

All isotypes

Uni-Gold™ HIV-1/2



10 min

HIV-1 gp41, gp120 RP
HIV-2 p36 RP

IgG

Introduction

HIV diagnosis in resource–restricted countries depend on rapid diagnostic tests (RDTs)

The HIV diagnostic algorithm used in Tanzania
Sequential testing using two RDTs

1. SD Biotec HIV1/2



Reactivity in the 1st RDT
is confirmed by a 2nd RDT

2. Uni-Gold™ HIV-1/2

Reactive
2 lines of any intensity appear in
both the control and test areas.



Linkage to
treatment
and care



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Introduction

The HIV diagnostic algorithm used in Mozambique Sequential testing using two RDTs

1. Alere Determine HIV1/2

Reactive
2 lines of any intensity appear in both the control and patient areas.



Reactivity in the 1st RDT
is confirmed by a 2nd RDT

2. Uni-Gold™ HIV

Reactive
2 lines of any intensity appear in both the control and test areas.



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Objective

- To explore the impact of VISR on the performance of HIV rapid diagnostic tests and to evaluate two African countries' HIV diagnostic algorithms

Material and methods

Samples collected at peak immunogenicity

- 137 stored plasma/serum samples from healthy HIVIS/TaMoVac vaccinees collected 1 month after the final vaccination (collected 2009 to 2014)

Samples collected over time

Stored serum samples from healthy HIVIS03/06 vaccinees

- 29 samples collected 1 month after 3XHIV-DNA+2XHIV-MVA
- 23 samples collected 16 months after 3XHIV-DNA+2XHIV-MVA
- 20 samples collected 3 years after 3XHIV-DNA+2XHIV-MVA

Material and methods

ELISA (4th generation diagnostic assay)

- Enzygnost[®] HIV Integral 4 ELISA (Siemens Healthcare Diagnostics Products GmbH, Marburg, Germany)

Western blot

- MP Diagnostics[™] HIV Blot 2.2 western blot assay (Eschwege, Germany)

Rapid diagnostic tests

- SD Bioline HIV 1/2 3.0 (Standard Diagnostic Inc, Giheung-gu, Republic of Korea)
- Determine HIV 1/2 (Alere Medical Co.Ltd, Chiba, Japan)
- UniGold HIV (Trinity Biotech, Bray, Ireland)

ELISA (in-house vaccine-specific assay)

- Subtype C rgp140 (reported as end-point titer)

Results

VISR was common at peak immunogenicity

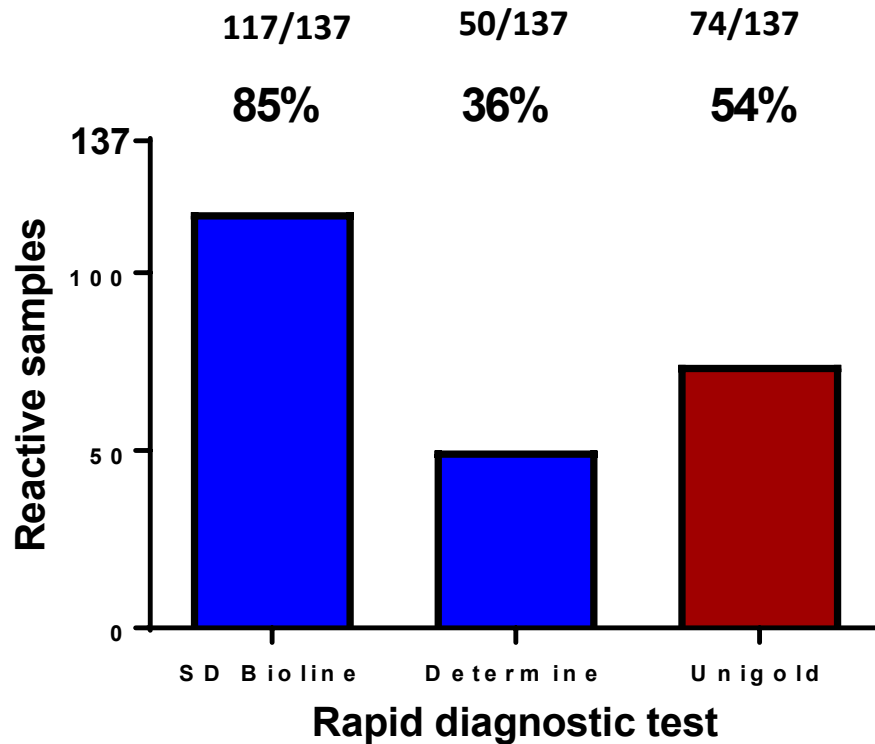
Reactivity in **Enzygnost[®] HIV Integral 4 ELISA**: 128/137 (93%)

Western Blot

Organization	Criteria for positive interpretation	Frequency of VISR
CDC and APHL	Presence of any two of p24, gp41, gp120/gp160 bands	136/137 (99.3%)
WHO	Presence of two ENV bands with or without GAG or POL	91/137 (66.4%)

Results

VISR was common at peak immunogenicity



Results

Missclassification by HIV diagnostic algorithm

Country	Algorithm	Rate of missclassification
Tanzania	Sequential testing SD Bioline HIV1/2, Uni-Gold™ HIV	74/137 (54%)
Mozambique	Sequential testing Alere Determine HIV1/2, Uni-Gold™ HIV	36/137 (26%)*

*Significantly lower, $p < 0.0001$

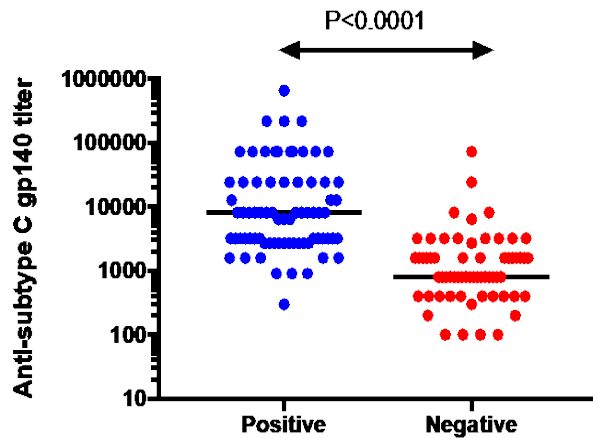
Results- antibody titers matter

(peak immunogenicity, n=137)

Tanzanian algorithm

SD Bioline HIV1/2 (screening)

UniGold™ HIV (confirmation)

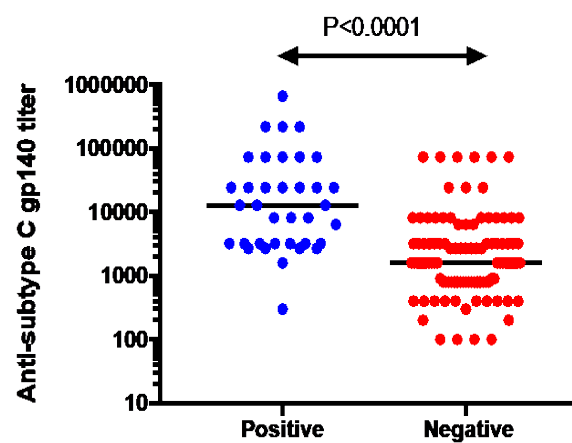


Classification

Mozambican algorithm

Determine HIV1/2 (screening)

UniGold™ HIV (confirmation)



Classification



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Results

The rate of missclassification wained over time in HIVIS03/06 vaccines (3xHIV-DNA+2XHIV-MVA)

Algorithm	Number of Reactive/ Number of Tested, (%)		
	1 month after the second HIV-MVA vaccination	16 months after the second HIV-MVA vaccination	3 years after the second HIV-MVA vaccination
Tanzanian	14/29 (48)	2/23 (8.7)	0/20
Mozambican	7/29 (24)	2/23 (8.7)	0/20

Conclusion

- The HIV diagnostic algorithms used in Tanzania and Mozambique will misclassify a substantial proportion of healthy HIV vaccine recipients
- Efforts are needed to develop simple, affordable, serological or molecular tools that can discriminate VISR from true HIV infection at the point of care.

Msafiri F et al. *Vaccines* **2022**, 10, 1062; doi. 10.3390/vaccines10071062

The volunteers

FoHM/KI

Gunnel Biberfeld
Britta Wahren
Charlotta Nilsson
Karina Godoy
Lindvi Gudmundsdotter
Gunnel Engström
Andreas Bråve
Karl Ljungberg
Karolinska/SöS
Eric Sandström
Bo Hejdeman



Örebro University

Sören Andersson

INS/CISPOC-Mozambique

Ilesh Jani
Edna Viegas
Nelson Tembe
Bindiya Meggi
Nafissa Osman
NIMR-Tanzania
Sayoki Mfinanga
Mbazi Senkoro



MUHAS-Tanzania

Fred Mhalu
Muhammad Bakari
Eligius Lyamuya
Said Aboud
Agricola Joachim
Patricia Munseri
Deus Buma
Candida Moshiro
Eric Aris
Mohamed Janabi
Kisali Pallangyo
Edith Tarimo



MMRC-Tanzania

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Asli Bauer

Imperial College London

Frances Gotch
Roger Tatoud

MRC; UK

Sheena McCormack
Sarah Joseph
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Mary Marovich
Jeffrey Currier
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Nelson Michael



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Thank you!

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